

# *Signaling pathways and cell cycle: modeling control mechanisms*

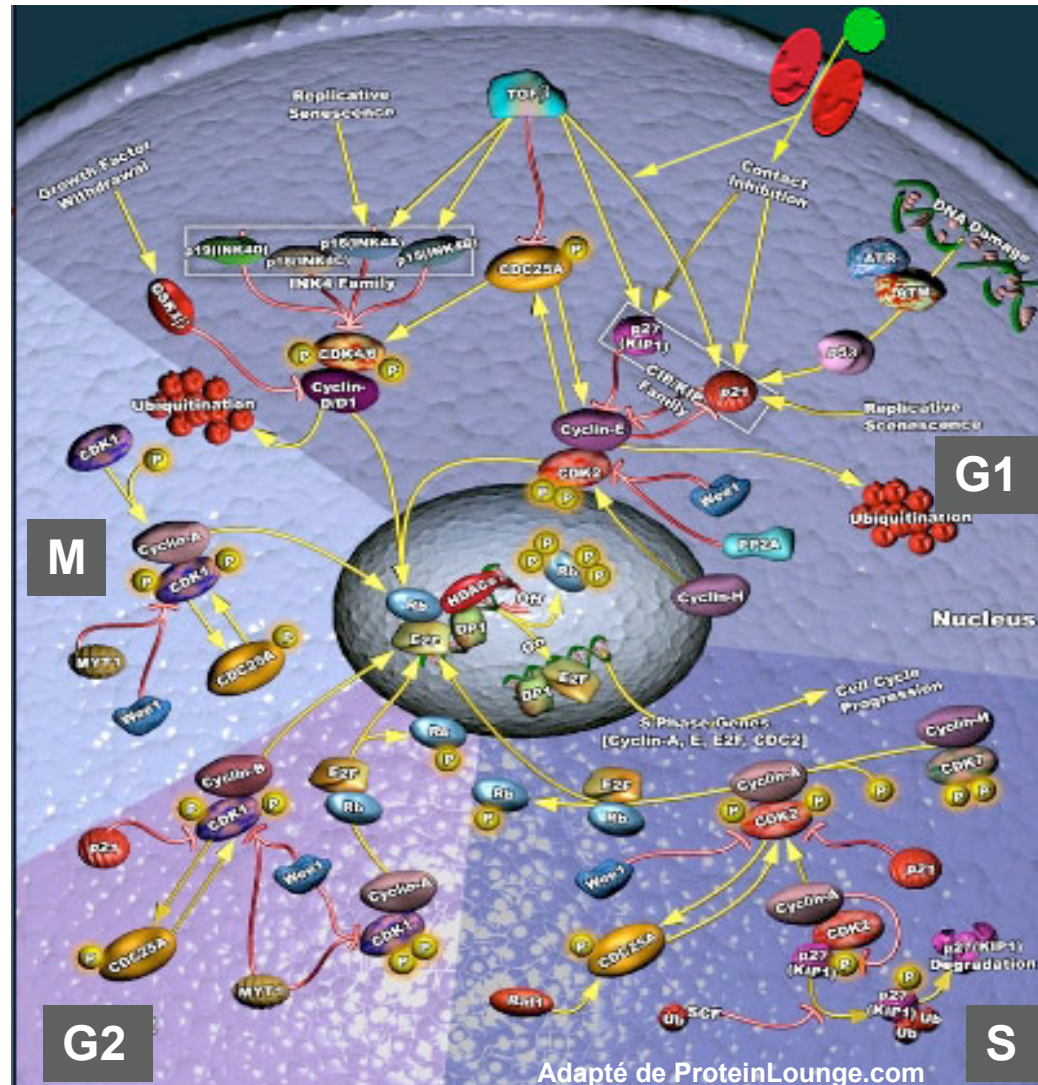
*Nolwenn Le Meur*

*J. Gruel - M. Le Borgne - N. Théret*

*6<sup>ème</sup> journée de la plate-forme GenOuest*

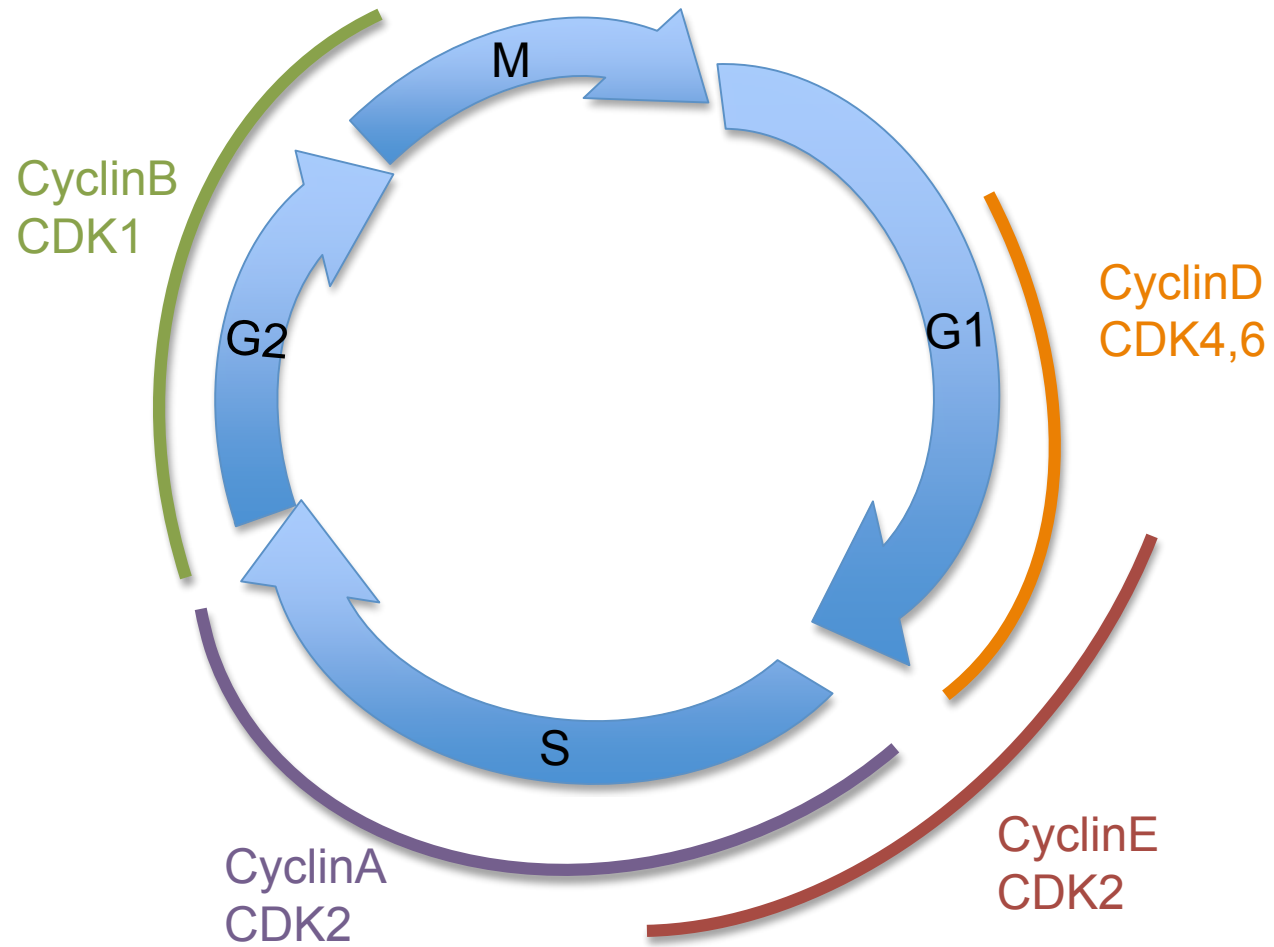
*Rennes, 21 October 2008*

# The role of signaling pathways in controlling cell cycle

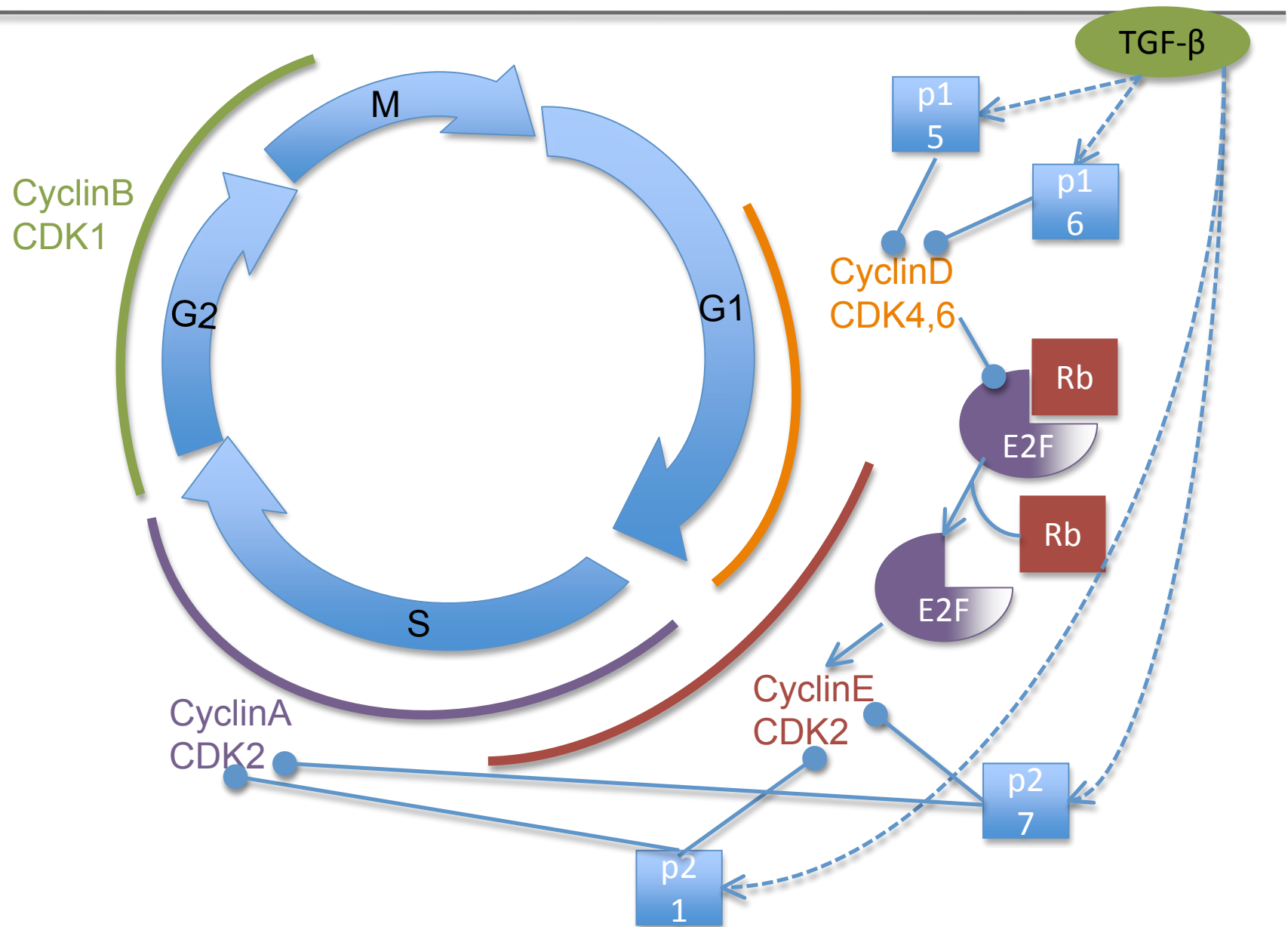


# Cell cycle [Basic]

---



# Cell cycle and signaling pathway [Basic]

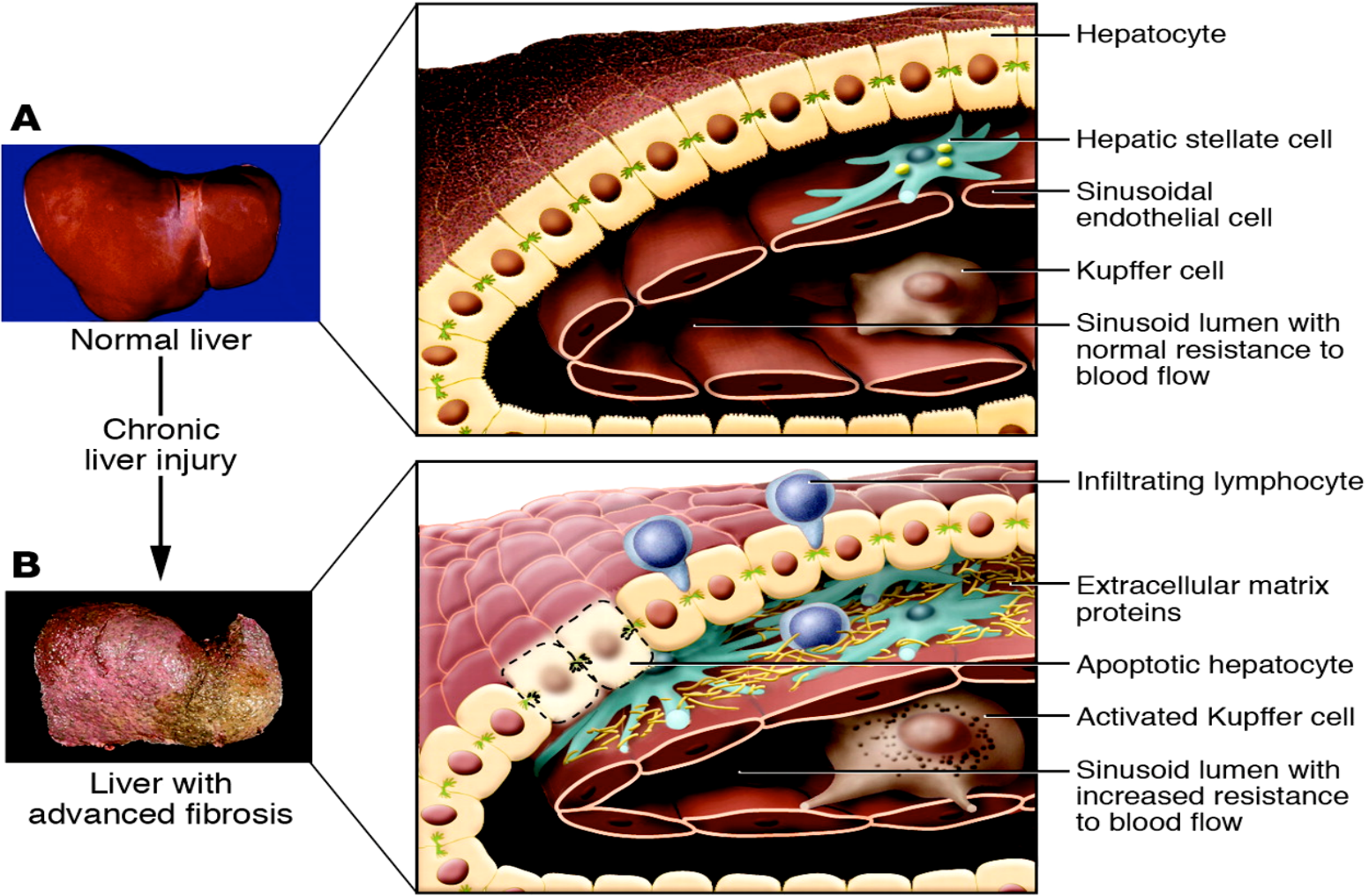


# Cell cycle, Signaling and Cancer

Name of gene	Nature of protein function	Type of tumor
<i>RARβ2</i>	nuclear receptor for differentiation	breast, lung
<i>p57<sup>Kip2</sup></i>	CDK inhibitor	gastric, pancreatic, hepatic; AML
<i>TIMP3</i>	inhibitor of metalloproteinases	diverse tumors
<i>IGFBP</i>	sequesters IGF-1 factor	diverse tumors
<i>CDKN2A/p16<sup>INK4A</sup></i>	inhibitor of CDK4/6	diverse tumors
<i>CDKN2B/p15<sup>INK4B</sup></i>	inhibitor of CDK4/6	diverse tumors
<i>p14<sup>ARF</sup></i>	inhibitor of HDM2/MDM2	colon, lymphoma
<i>APC</i>	inducer of β-catenin degradation	colon carcinomas
<i>p73</i>	aids p53 to trigger apoptosis	diverse tumors
<i>GSTP1</i>	mutagen inactivator	breast, liver, prostate
<i>MGMT</i>	DNA repair enzyme	colorectal
<i>CDH1</i>	cell-cell adhesion receptor	bladder, breast, colon, gastric
<i>DAPK</i>	kinase involved in cell death	bladder
<i>MLH1</i>	DNA mismatch repair enzyme	colon, endometrial, gastric
<i>TGFBR2</i>	TGF-β receptor	colon, gastric, small-cell lung
<i>THBS1</i>	angiogenesis inhibitor	colon, glioblastoma
<i>RB</i>	cell-cycle regulator	retinoblastoma
<i>CASP8</i>	apoptotic caspase	neuroblastoma, SCLC
<i>APAF1</i>	pro-apoptotic cascade	melanoma
<i>CTMP</i>	inhibitor of Akt/PKB	glioblastoma multiforme

Weinberg (2006) *The biology of cancer*. Garland

# From fibrosis to hepatocellular carcinoma



Battler R. and Brenner D.A (2007). *J. Clin. Inv.*

# TGF- $\beta$ signaling pathway

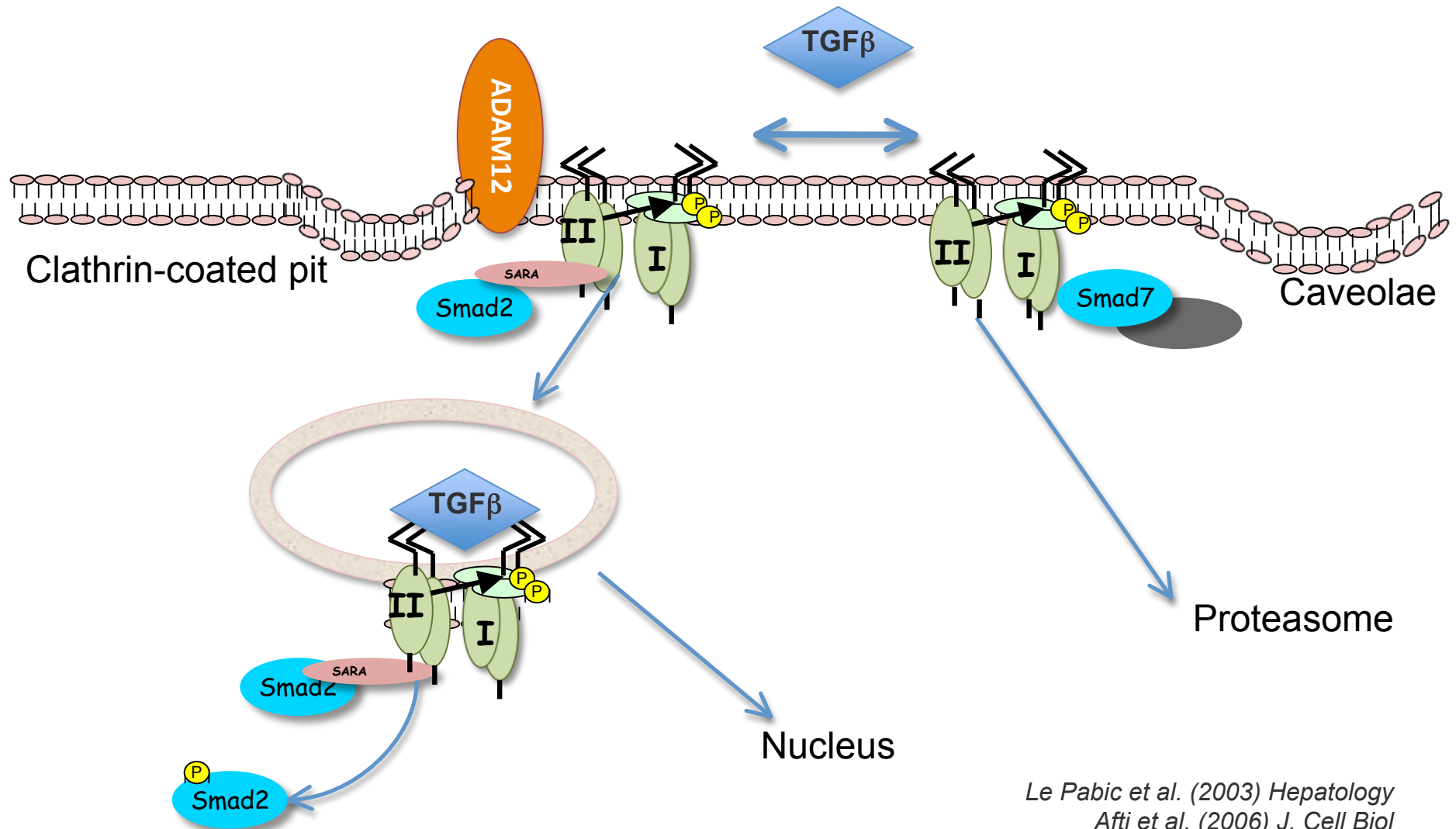
---

- Extracellular matrix production
- Cell proliferation
- Functional differentiation
- Cell motility
- Apoptosis

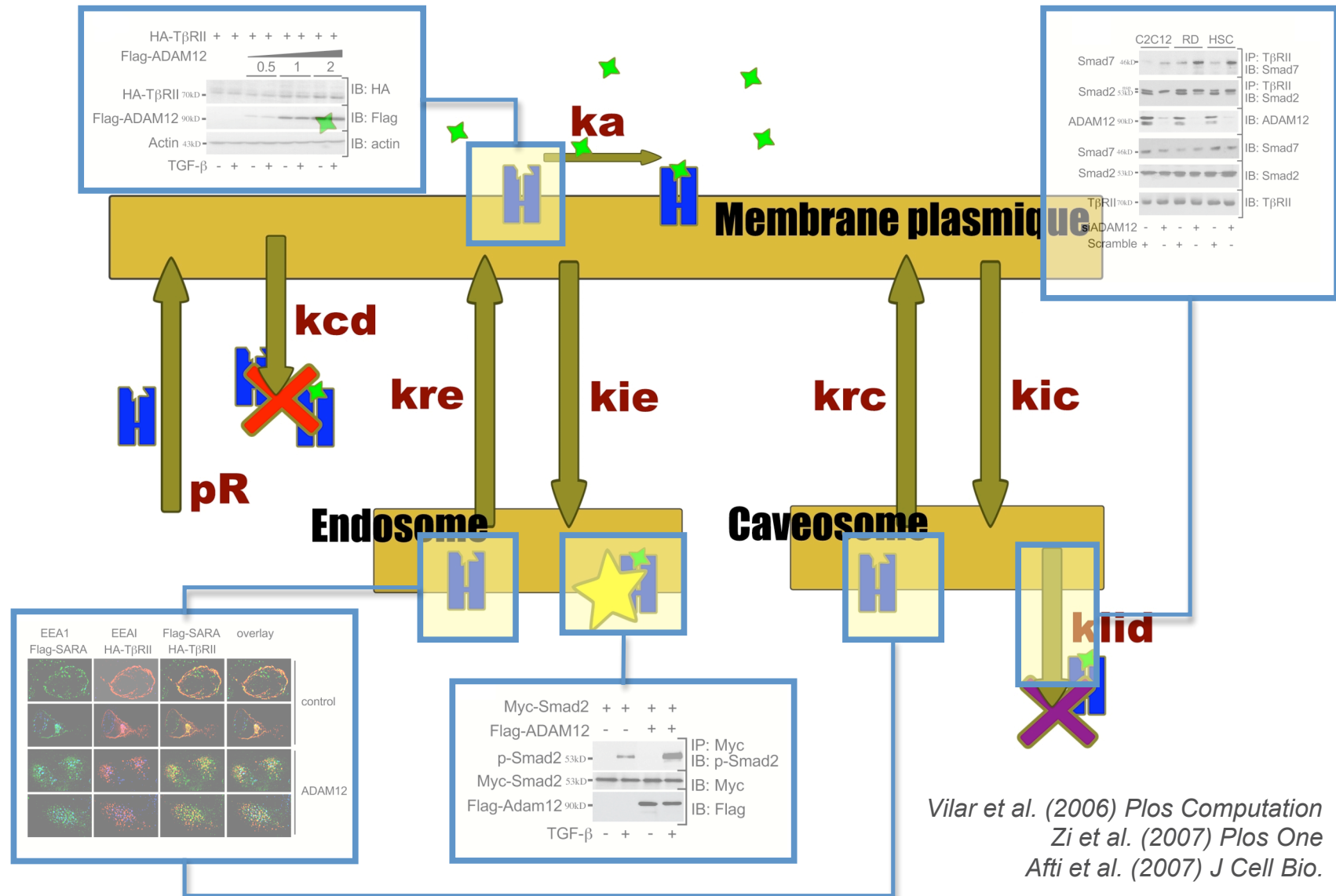
# ADAM12 and TGF- $\beta$ receptors



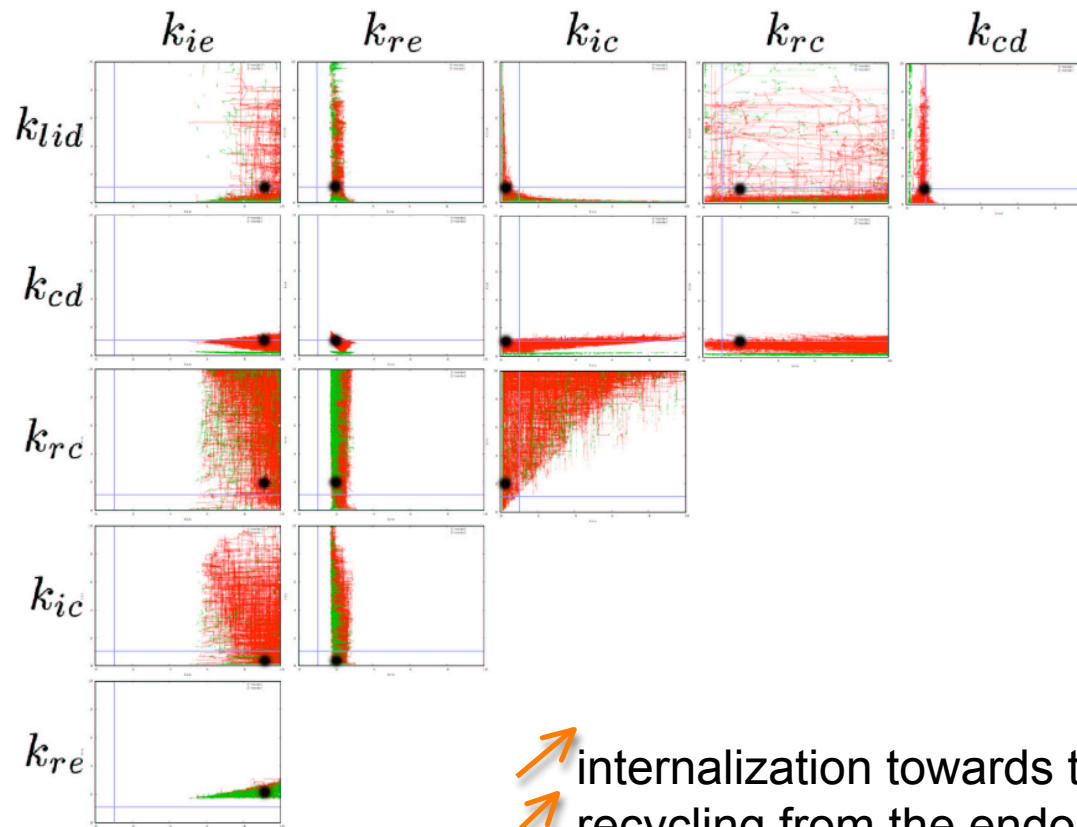
# TGF- $\beta$ signaling pathways



# ADAM12 in TGF- $\beta$ receptor trafficking

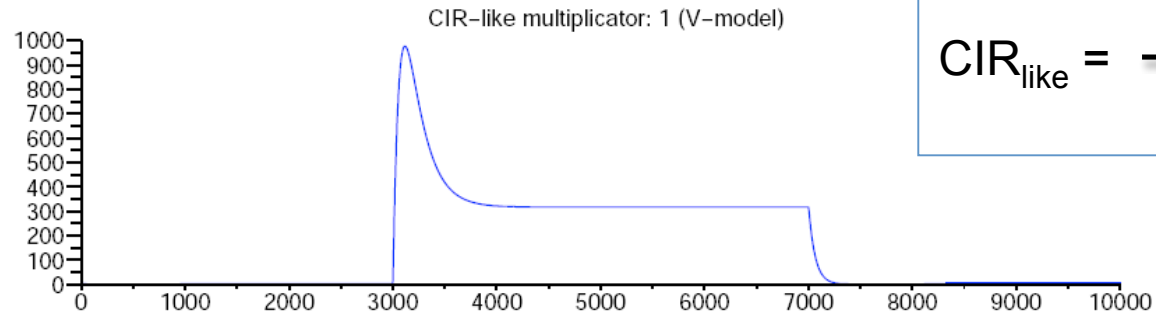


# ADAM12 and TGF- $\beta$ signal strength

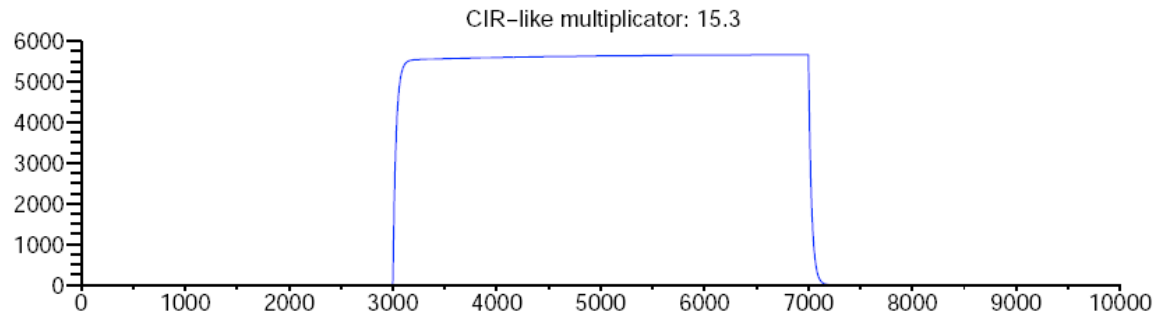
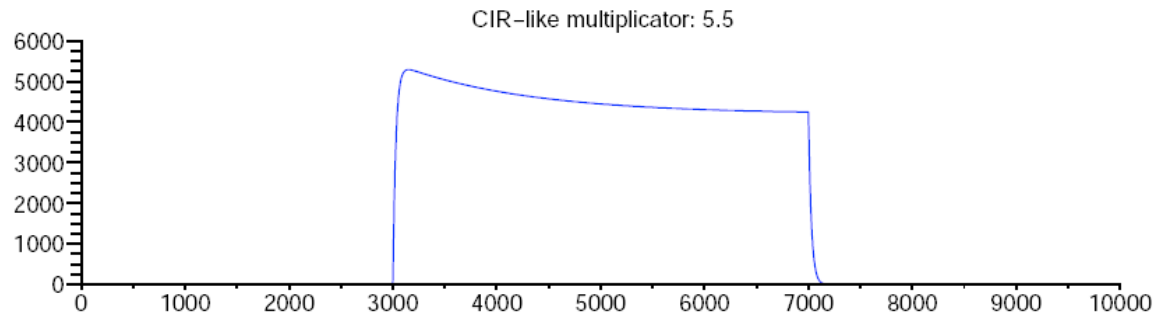


- ↗ internalization towards the endosome
- ↖ recycling from the endosome
- ↓ constitutive degradation

# ADAM12 and TGF- $\beta$ signal shape



$$\text{CIR}_{\text{like}} = \frac{\text{Constitutive}}{\text{Ligand-induced}}$$



TGF- $\beta$   
signaling  
activity

Time

*J. Gruel et al in preparation*

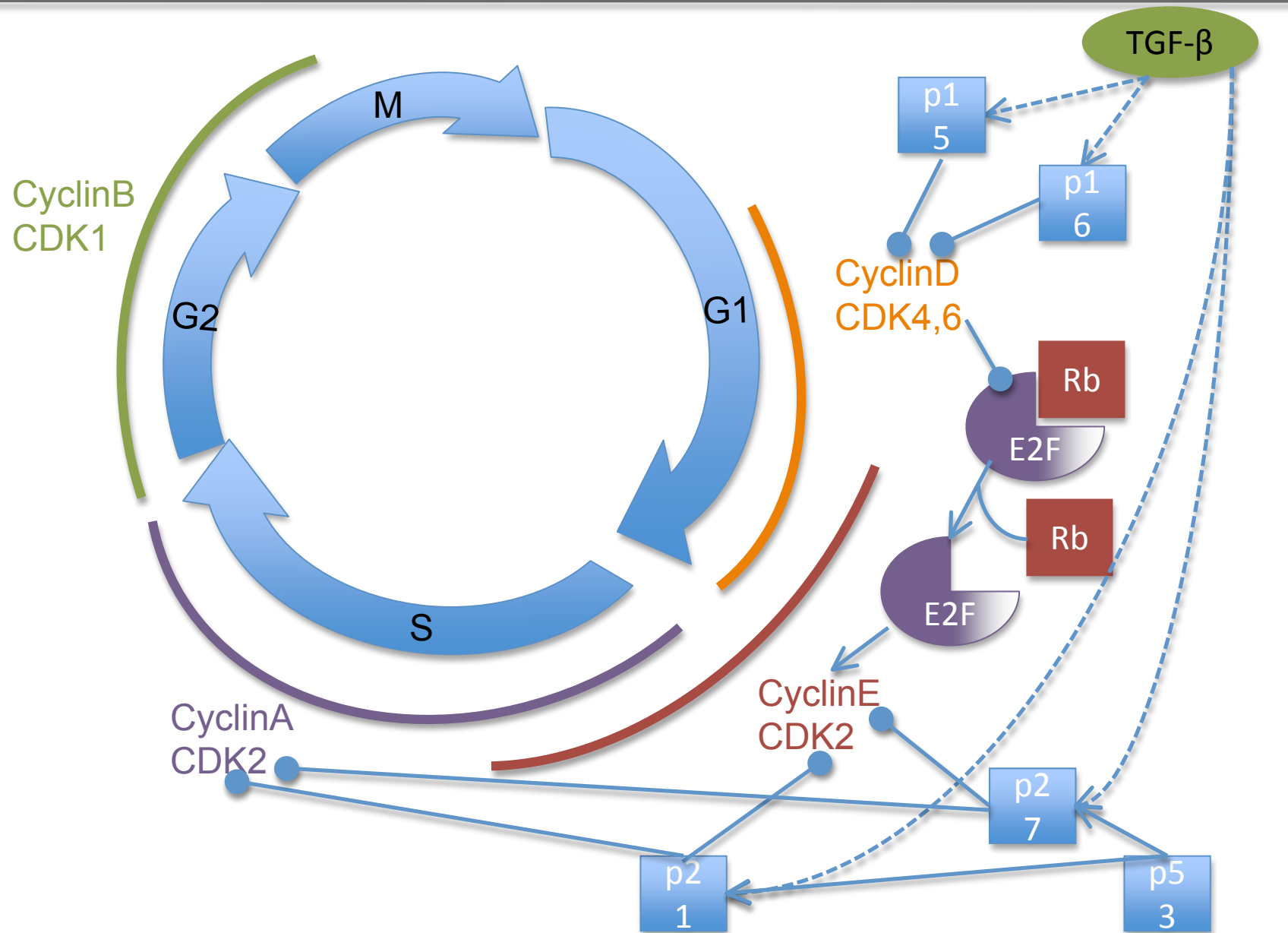
## Fitting algorithm as a exploratory tool

---

- Mathematical model
  - Differential equation
  - Solution space
- Biological insights
  - Signal strength
  - Signal shape

# Signaling pathways and cell cycle

# Cell cycle and signaling pathway [Basic]



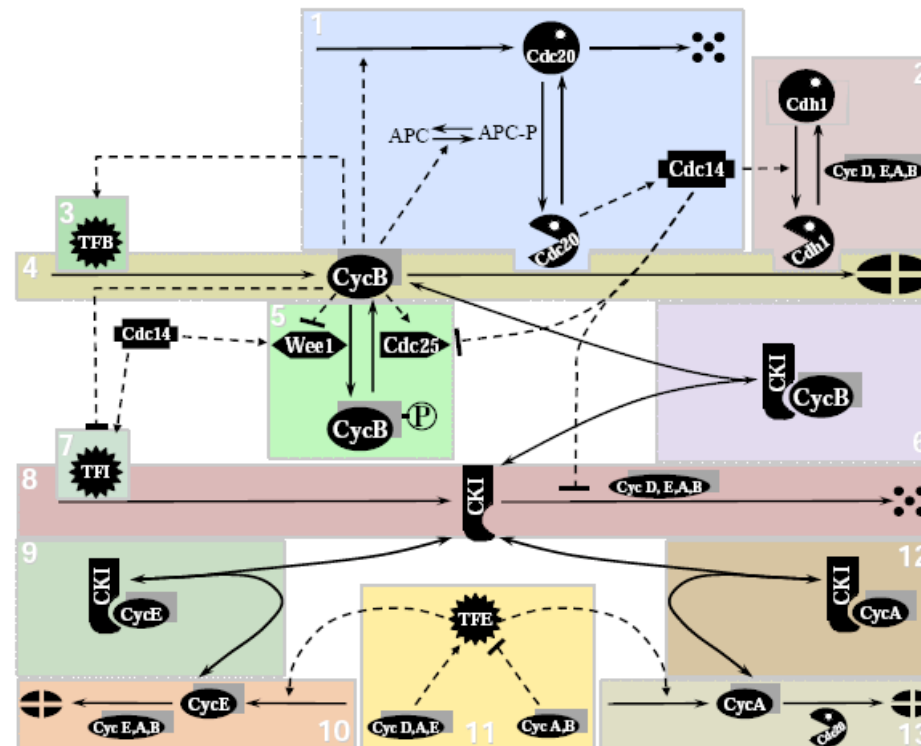
# Modeling the Cell Cycle

Biophysical Journal Volume 90 June 2006 4361–4379

## Analysis of a Generic Model of Eukaryotic Cell-Cycle Regulation

Attila Csikász-Nagy,<sup>\*†</sup> Dorjsuren Battogtokh,<sup>\*</sup> Katherine C. Chen,<sup>\*</sup> Béla Novák,<sup>†</sup> and John J. Tyson<sup>\*</sup>

<sup>\*</sup>Department of Biological Sciences, Virginia Polytechnic Institute and State University, Blacksburg, Virginia 24061-0406; and <sup>†</sup>Molecular Network Dynamics Research Group of the Hungarian Academy of Sciences and Department of Agricultural and Chemical Technology, Budapest University of Technology and Economics, H-1521 Budapest, Hungary





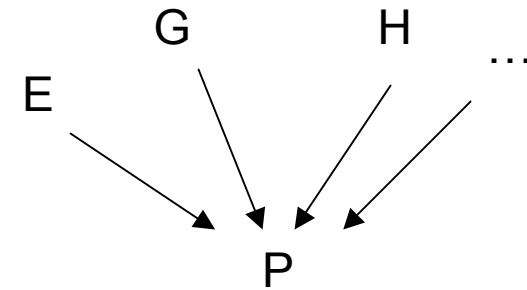
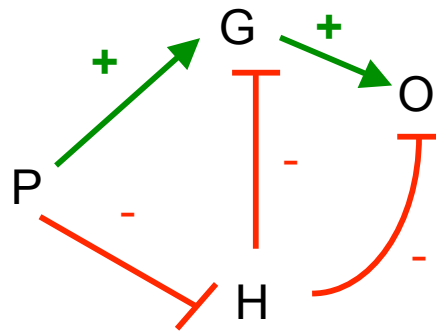
# Network Modeling methods

---

*Qualitative*

*Quantitative*

*Dynamic*



$$dP/dt = K_{ev}(E) + K_{gv}(G) + \dots$$

# Modeling the Cell Cycle

---

OPEN ACCESS Freely available online



## Boolean Network Model Predicts Cell Cycle Sequence of Fission Yeast

**Maria I. Davidich, Stefan Bornholdt\***

Institut für Theoretische Physik, Universität Bremen, Bremen, Germany

**BIOINFORMATICS**

Vol. 22 no. 14 2006, pages e124–e131  
doi:10.1093/bioinformatics/btl210

---

## Dynamical analysis of a generic Boolean model for the control of the mammalian cell cycle

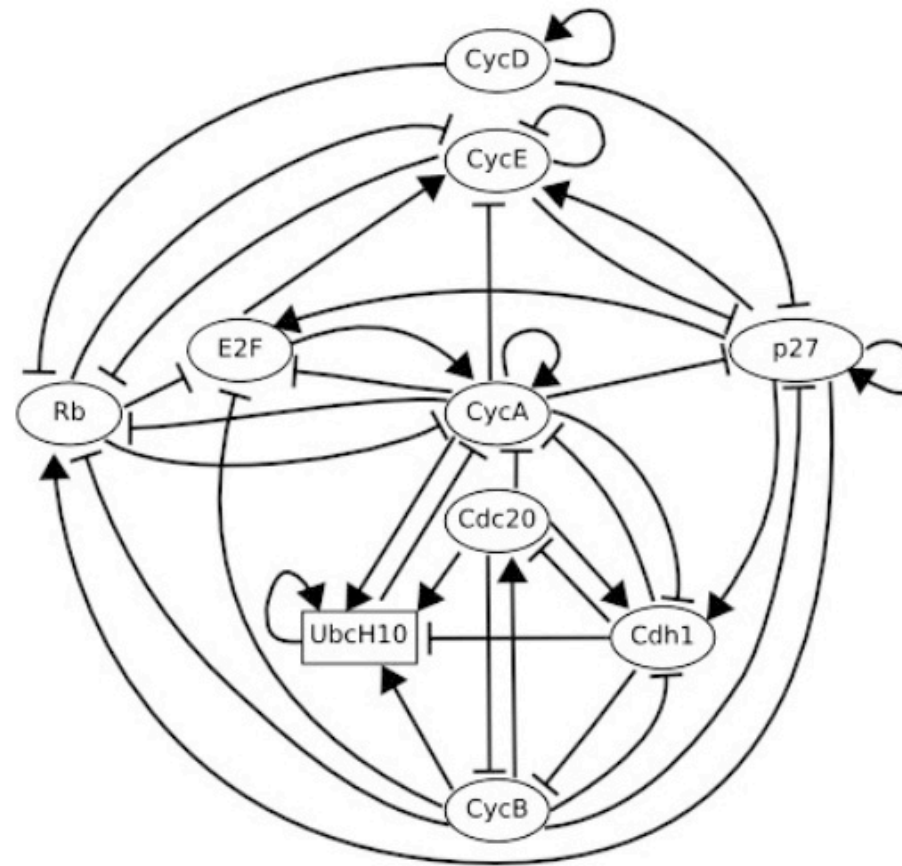
Adrien Fauré, Aurélien Naldi, Claudine Chaouiya and Denis Thieffry\*

Institut de Biologie du Développement de Marseille-Luminy, Campus scientifique de Luminy, CNRS case 907, 13288 Marseille, France

---

# Regulatory Graph of Cell Cycle

---



**Fig. 1.** Logical regulatory graph for the mammalian cell cycle network. Each node represents the activity of a key regulatory element, whereas the edges represent cross-regulations. Blunt arrows stand for inhibitory effects, normal arrows for activations.

*Fauré et al. (2006) Bioinformatics*

# Logical States

---

Regulatory graph  $G = \{g_1, \dots, g_n\}$

Logical state vector  $S = (s_1, \dots, s_n)$   $s_i \in \{0, \dots, Max_i\}$

CycD	Rb	E2F	CycE	CycA	P27	Cdc20	Chd1	UbcH10	CycB
1	0	1	1	0	0	0	1	0	0

# Temporal evolution in the cell cycle

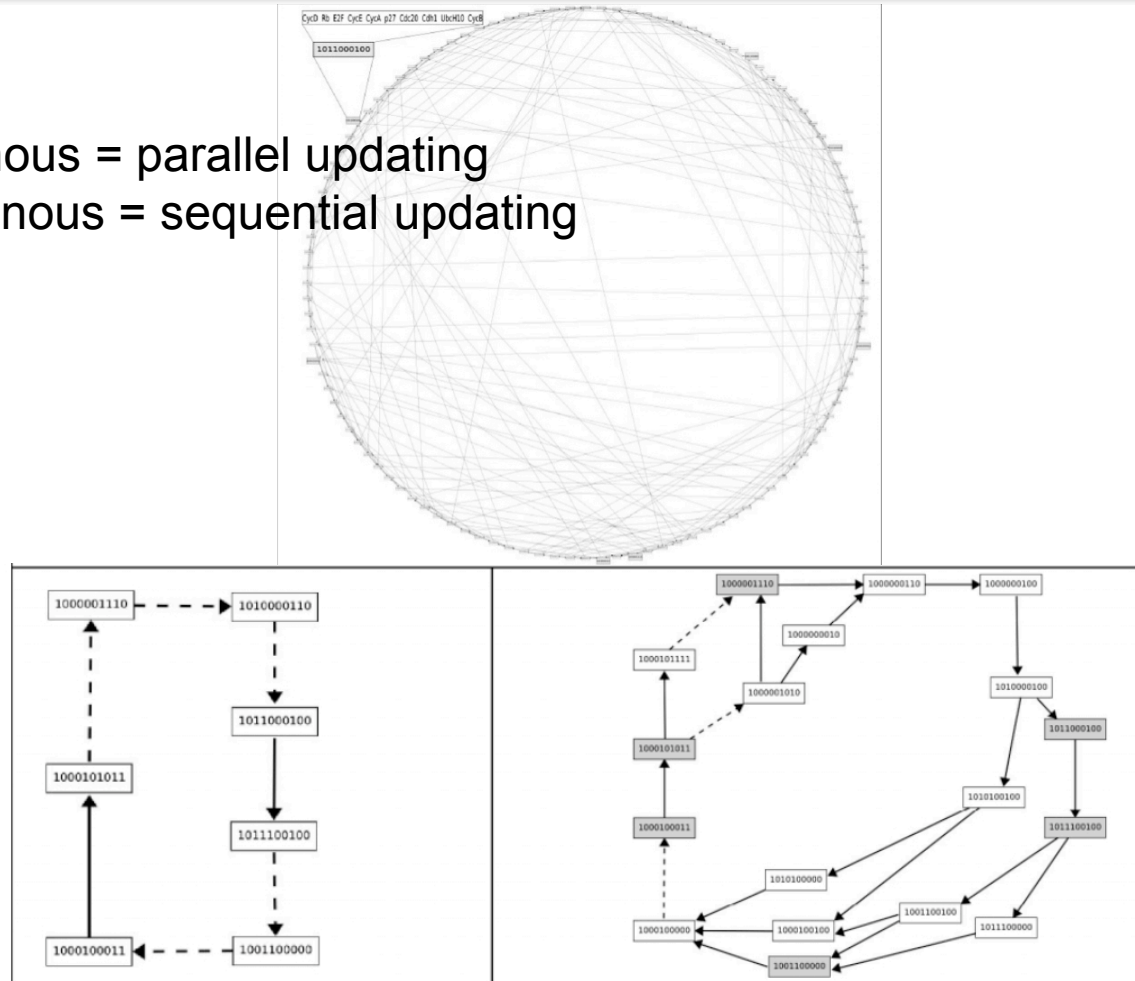
**Table 2.** Temporal evolution of protein states in the cell cycle network.

Time Step	Start	SK	Cdc2/Cdc13	Ste9	Rum1	Slp1	Cdc2/Cdc13*	Wee1 Mik1	Cdc25	PP	Phase	comments
1	1	0	0	1	1	0	0	1	0	0	START	Cdc2/Cdc13 dimers are inhibited, antagonists are active.
2	0	1	0	1	1	0	0	1	0	0	G1	SK are becoming active
3	0	0	0	0	0	0	0	1	0	0	G1/S	When Cdc2/Cdc13 and SK dimers switch off Rum1 and Ste9/APC, the cell passes 'Start' and DNA replication takes place, Cdc2/Cdc13 starts to accumulate
4	0	0	1	0	0	0	0	1	0	0	G2	Activity of Cdc2/Cdc13 achieves moderate level, which is enough for entering G2 phase but not mitosis, since Wee1/Mik1 inhibits the activity of residue Tyr-15 of Cdc2 (Cdc2/Cdc13* is not active)
5	0	0	1	0	0	0	0	0	1	0	G2	Moderate activity Cdc2/Cdc13 activates Cdc25
6	0	0	1	0	0	0	1	0	1	0	G2/M	Cdc25 reverses phosphorylation, removing the inhibiting phosphate group and activating Cdc2/Cdc13*
7	0	0	1	0	0	1	1	0	1	0	G2/M	Cdc2/Cdc13* reaches high activity level sufficient to activate Slp1/APC mitosis
8	0	0	0	0	0	1	0	0	1	1	M	Slp1 degrades Cdc13, that is inhibits complex Cdc2/Cdc13 and Cdc2/Cdc13*.
9	0	0	0	1	1	0	0	1	0	1	M	Antagonists of Cdc2/Cdc13 are reset.
10	0	0	0	1	1	0	0	1	0	0	G1	Cell reaches G1 stationary state (PP is inactive)

*Davidich et al. (2006) Plos One*

# Synchronous versus Asynchronous

Synchronous = parallel updating  
 Asynchronous = sequential updating



**Fig. 2.** Simulations of the wild-type cell cycle based on the Boolean model defined in Figure 1 and Table 1. Each vertex (node) represents one state, with the regulatory components ordered as mentioned in the top panel. The three state transition graphs correspond to the comprehensive asynchronous (top), the synchronous (bottom left), and a mixed (bottom right) assumptions. Note the difference of complexity between the asynchronous and synchronous graphs. In the bottom panels, solid arrows stand for single transitions, and dotted arrows for multiple transitions. The seven states involved in the synchronous cycle are grey-shaded in the asynchronous and mixed state transition graphs. For larger resolution pictures, see *GINsim* website.

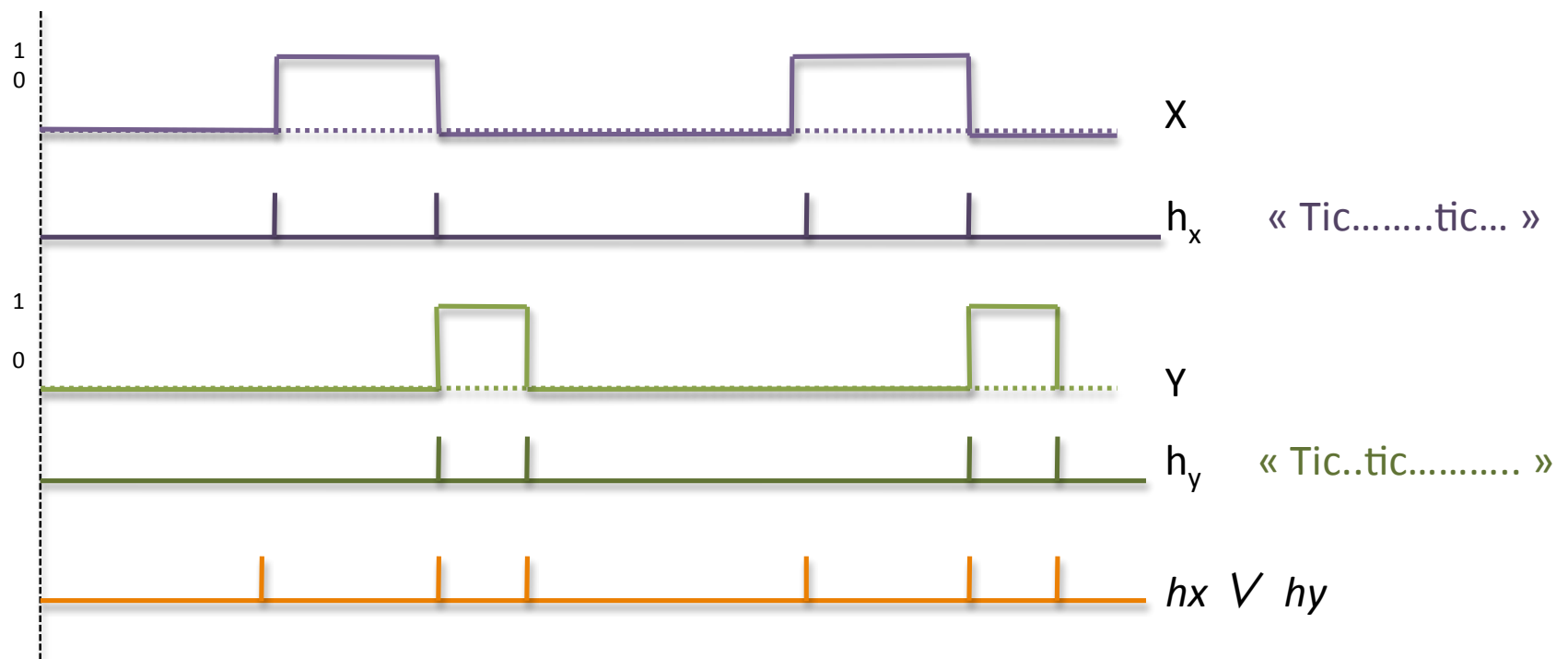
# Discrete dynamic modeling

---

- Signal updating in the model
  - Synchronous
  - Asynchronous
  - Interlacing
- Answer questions
  - Is molecule A always before molecule B?
  - Are C and E never active at the same time?
  - ...

# Discrete multi-clock modeling: a new formalism

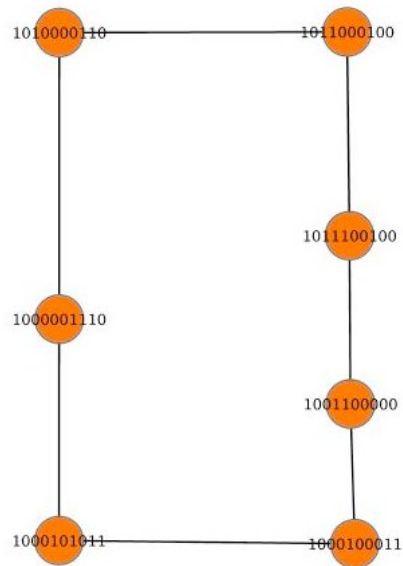
- Time in the model
- State  $X$  is modified by a signal *with* a clock  $h_x$



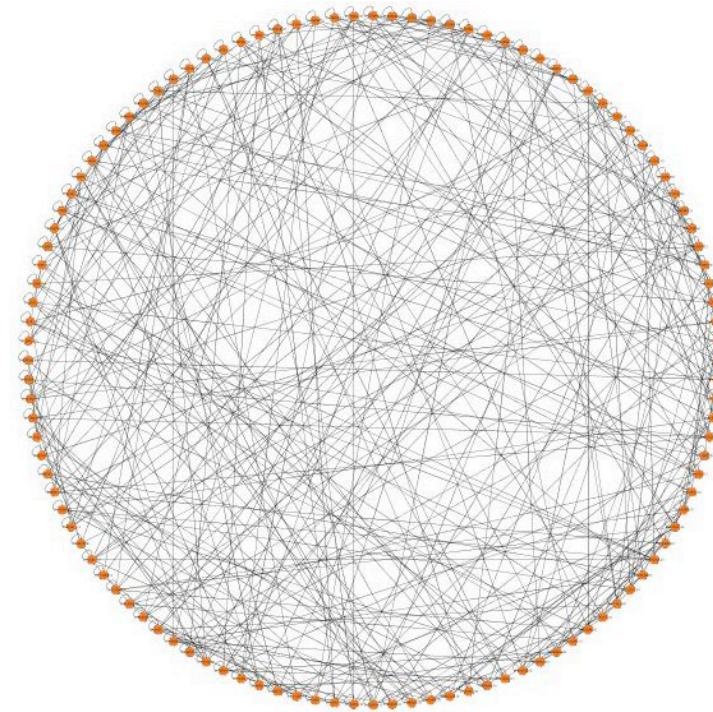


# Discrete multi-clock modeling: first results

State:  $D, f(), D0$   
Clock:  $h\_D$   
 $F() = A$  and  $B$  when  $h\_D$



synchronous



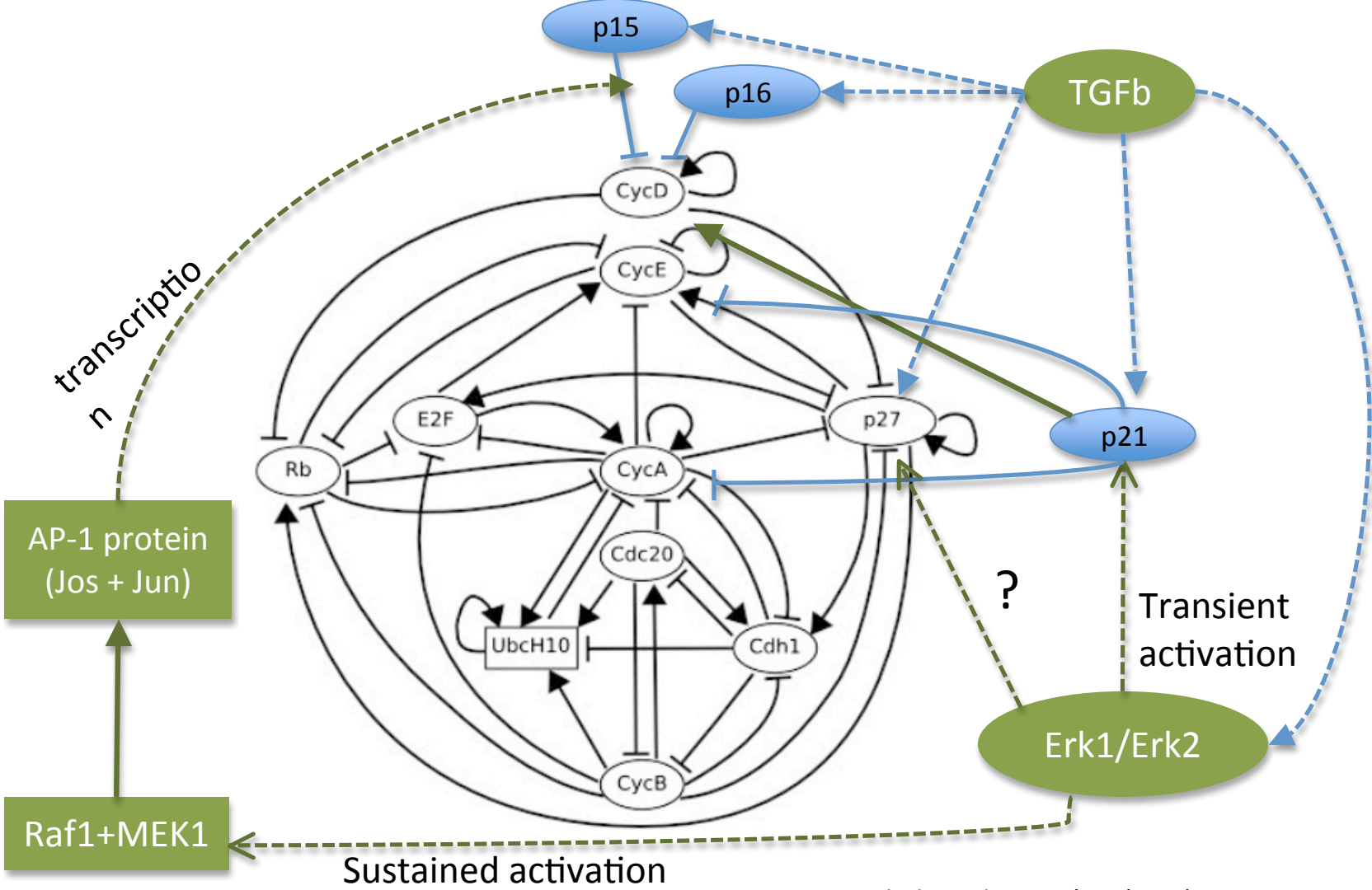
asynchrone

## Future work

---

- Temporal constraints
- Methods and metrics
  - analysis
  - diagnostics
- Validation
  - *In silico*
  - *In vivo*
- Interface biologist/computational model
- Include signaling pathways

# Plug-ins: Signaling Pathways



Meloche and Pouyssegur (2007) Oncogene

# Acknowledgements

---

- ✧ N. Théret
- ✧ G. Baffet
- ✧ Y. Arlot
- ✧ D. Lagadic-Gossmann

- ✧ M. Le Borgne
- ✧ J. Gruel
- ✧ O. Radulescu
- ✧ J. Nicolas



**INRIA**

**Inserm**

Institut national  
de la santé et de la recherche médicale





# Kinase and cell cycle

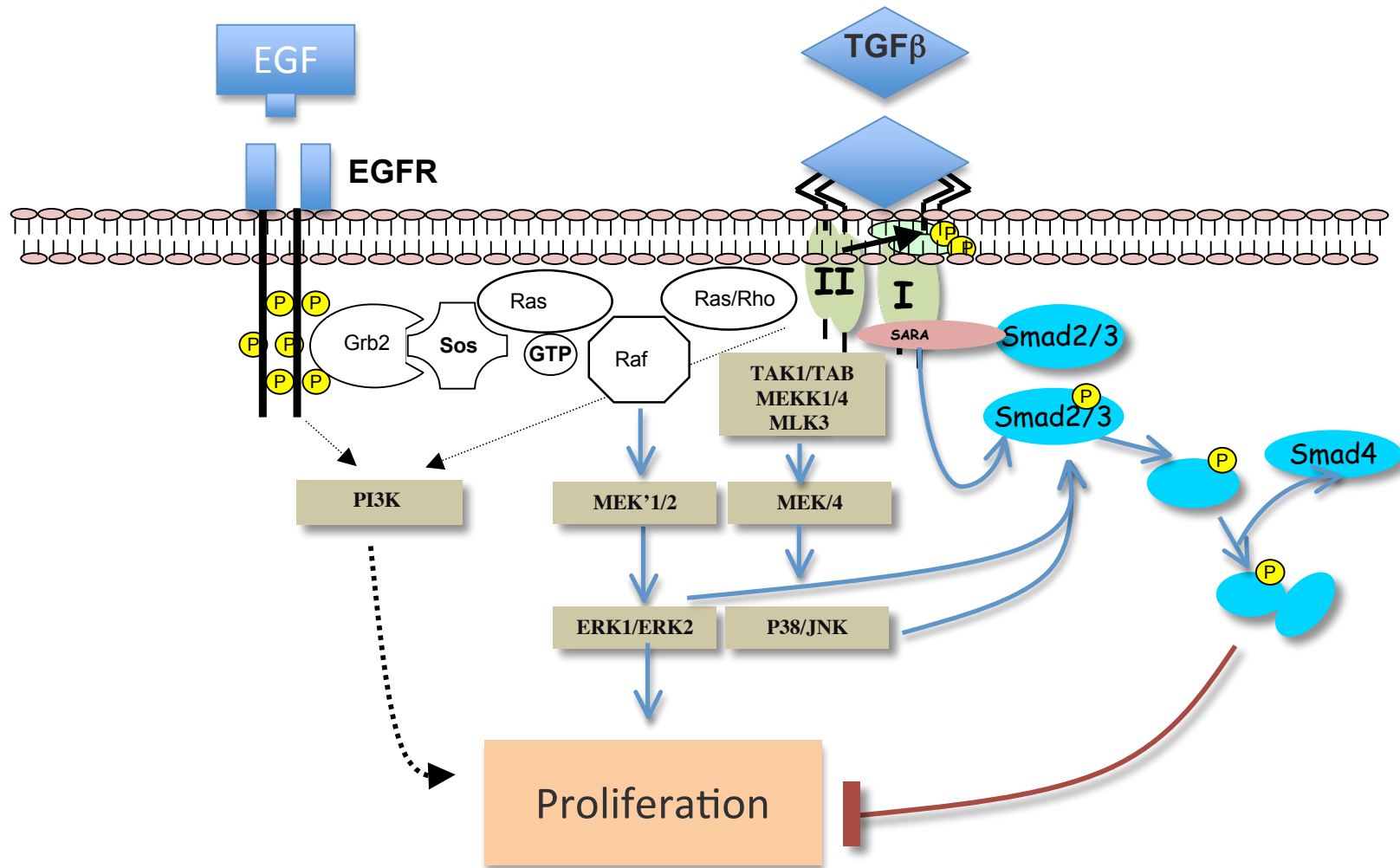
---

**Table 1.** Principles of cdk Inhibition

Inhibition of cdk4/6 leads to potent Rb-dependent G <sub>1</sub> arrest; however, compensation by cdk2 is possible
Inhibition of cdk4/6, cdk2, and cdk1 frequently results in arrest at the G <sub>1</sub> -S and G <sub>2</sub> -M boundaries
More selective inhibition of cdk2 and cdk1 leads to less potent G <sub>1</sub> arrest, S→G <sub>2</sub> effects, and E2F-1–dependent apoptosis
Recruitment to S phase by chemotherapy agents may sensitize cells to cdk inhibition; cdk inhibitor–mediated death may occur by E2F-1–dependent apoptosis, augmentation of DNA damage, or inhibition of DNA repair
Cdk1 inhibition may augment cell death after mitotic checkpoint activation by taxanes or KSP inhibitors
Highly selective cdk2 inhibition does not have antiproliferative effects in many cancer cell types
Inhibition of cdk9 preferentially depletes mRNAs with short half-lives (eg, Mcl-1, cyclin D1, c-myc, p53-induced p21 <sup>Waf1/Cip1</sup> , and hypoxia-induced VEGF)
Abbreviations: cdk, cyclin-dependent kinase; Rb, retinoblastoma protein; KSP, hsEg5 or kinesin-5; VEGF, vascular endothelial growth factor.

*Shapiro G. (2006) Journal of Clinical Oncology*

# EGF and TGF- $\beta$



# The role of signaling pathways in controlling cell cycle

