

# The POSEIDON Project: Protistian diversity as revealed by Ribosomal Barcoding along the TARA OCEANS' circumnavigation.

- Global Presentation of The Tara ocean's circumnavigation: Frederic Mahé
- Bioinformatics analyses of Barcode data: Stéphane Audic

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CNRS UPMC INSU

**Station Biologique  
Roscoff**

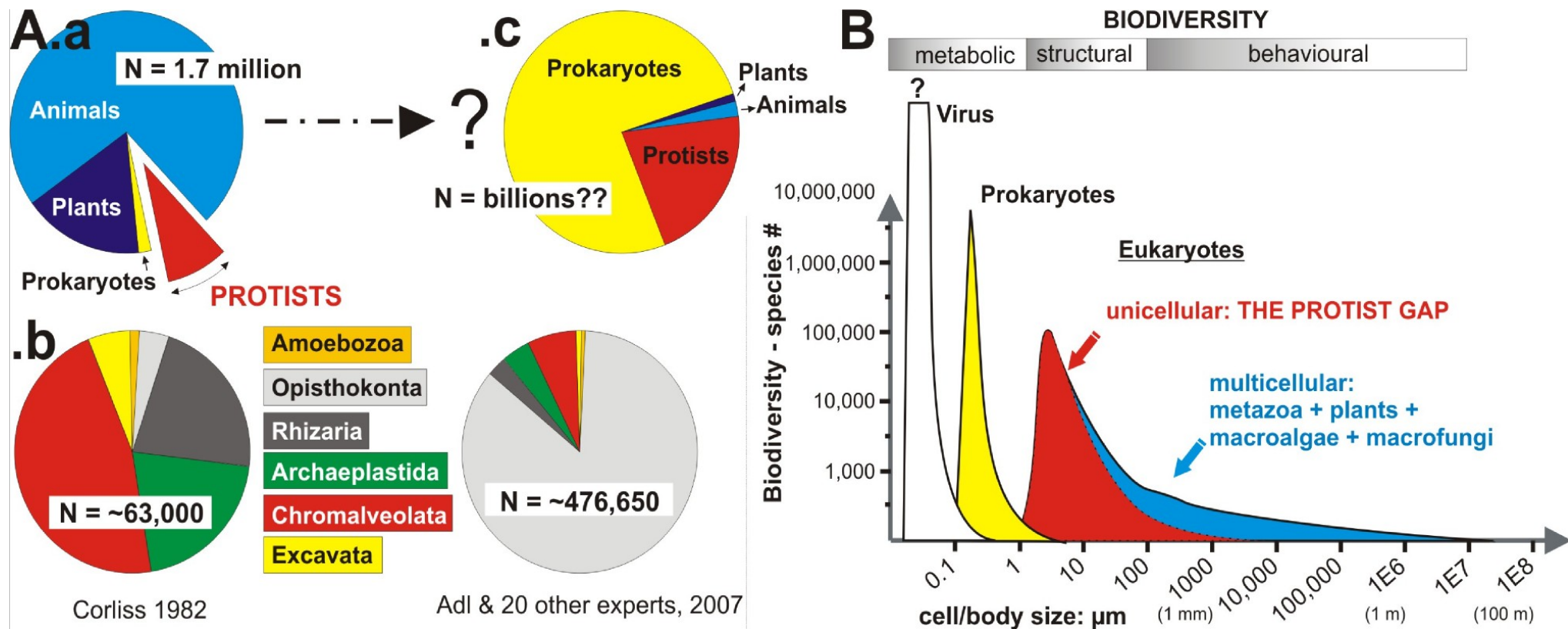


Figure 2: The marine protist biodiversity gap. A. a: Total number of living phenotypically described species on Earth ( $N=1.7$  million; from International Union for Conservation of Nature). The relatively lower numbers of unicellular (pro- and eu-karyotic) species principally reflects the history of biological science, dominated by de visu and microscopy observations since the 1600s. b: 1982 (single author) and 2007 (consortium) surveys of described protist species and their classification among the higher level protist super-groups. These censuses are highly dependant on the number (and character!) of experts working on particular groups. The 2007 picture is for instance largely skewed toward fungi and diatoms; estimation of the “real” biodiversity by the same expert consortium, based on preliminary molecular datasets and gut-feeling, reached  $\sim 18$  million species. Environmental (e)-rDNA clone library based studies over the last decade, have shown that eukaryotic microbial diversity is far greater than previously catalogued. Recent group-specific PCR of shorter e-DNA fragments (which reduces classical PCR biases) have further shown that protist biodiversity remains largely undersampled even using standard molecular approaches (Fig. 3). Overall, the molecular exploration of biodiversity tends to support the hypothetical distributions shown in Ac. and B. Hypothetical number of species (a species being defined as a distinct genome/phenotype couple at a given time) of viruses, prokaryotes, and eukaryotes in relation to cell/body size. The red area depicts the “protist gap”, where most eukaryotic species still remain to be discovered, despite their fundamental biogeochemical importance and their key position as a bridge between prokaryote and animal/plant biodiversities.

# Some key figures

- 1 one ml of water, you have approximately
  - 10 millions viruses
  - 1 million prokaryots
  - 0.1 millions eukaryots
- In Tara, each samples is approximately 100 liters
  - $10^{12}$  viruses
  - $10^{11}$  prokaryots
  - $10^{10}$  eukaryots

# DNA Barcoding for metagenomics

- Find a sequence region with very conserved parts and very variable parts: this is not a trivial problem, but this is a prerequisite because you want
  - To design primers that are very specific
  - But give you information ->variability
- Why not directly sequence all DNA? It depends of your problem.
  - If you just want to know who is here:
    - Most of it is of no use to you!
    - So you save at many levels: time/cost/..

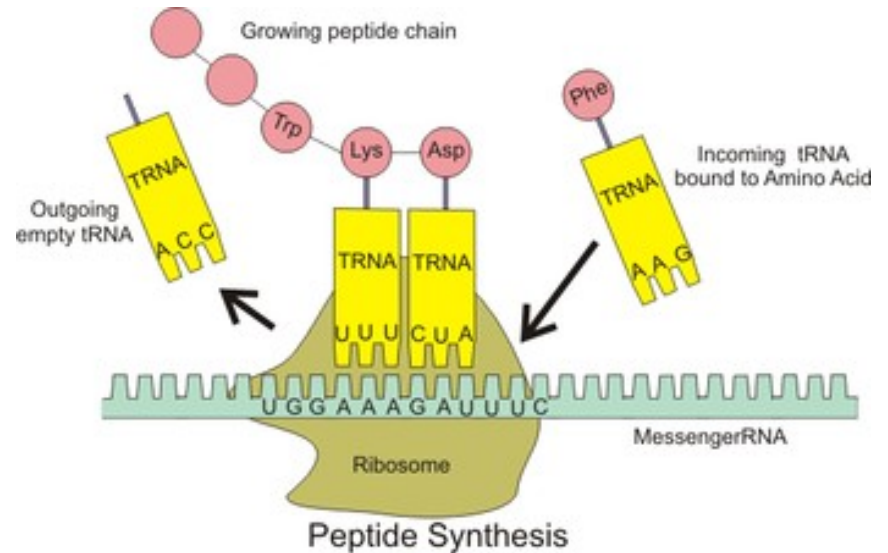
# What are good candidates for barcoding?

- It depends on what you are interested on:

- All living organisms have ribosomes with which they do protein assembly
- Among ribosome constituents ribosomal RNA is the more conserved between different species

- More targeted studies in general use other markers:

- See Barcode of Life project



# Many barcoding initiatives

**Barcode of Life**

Identifying Species with DNA Barcoding

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## Barcoding Projects

There are many international barcoding activities dedicated to the development of targeted public reference BARCODE sequence libraries.

FEATURED PROJECTS

Sort By:



### All Fungi Barcoding

All Fungi Barcoding provides up-to-date information on fungal barcoding and facilitates communication and collaboration among researchers interested in fungi.

[Project Site](#)



### Bee Barcode of Life Initiative (Bee-BOL)

Bee-BOL, the Bee Barcode of Life Initiative, is a global effort to coordinate the assembly of a standardized reference sequence library for all ~20,000 bee species. Bee-BOL is creating a valuable public resource in the form of an electronic database containing DNA barcodes, images, and geospatial coordinates of examined specimens. The database contains linkages to voucher specimens, information on species distributions, nomenclature, authoritative taxonomic information, collateral natural history information and literature citations.

[Project Site](#)



### Coral Reef Barcode of Life

The Coral Reef Barcode of Life campaign is a detailed barcode study of fishes at one site in the Great Barrier Reef to generate a barcode library that will aid taxonomic work by clarifying species boundaries and by revealing cryptic taxa.

[Project Site](#)



### European Consortium for the Barcode of Life (ECBOL)

ECBOL is an information and coordination hub on DNA barcoding in Europe organized within EDIT, the European Institute of Taxonomy and maintained by CBS, the Centraalbureau voor Schimmelmcultures in Utrecht. The ECBOL initiative (Calibrating European Biodiversity using DNA Barcodes) is a network of European researchers and is seeking to obtain funding from the coordination and maintenance of a Network of European Leading Labs.

[Project Site](#)

<http://www.barcodeoflife.org>

mitochondrial cytochrome c oxidase  
subunit I (CO1)

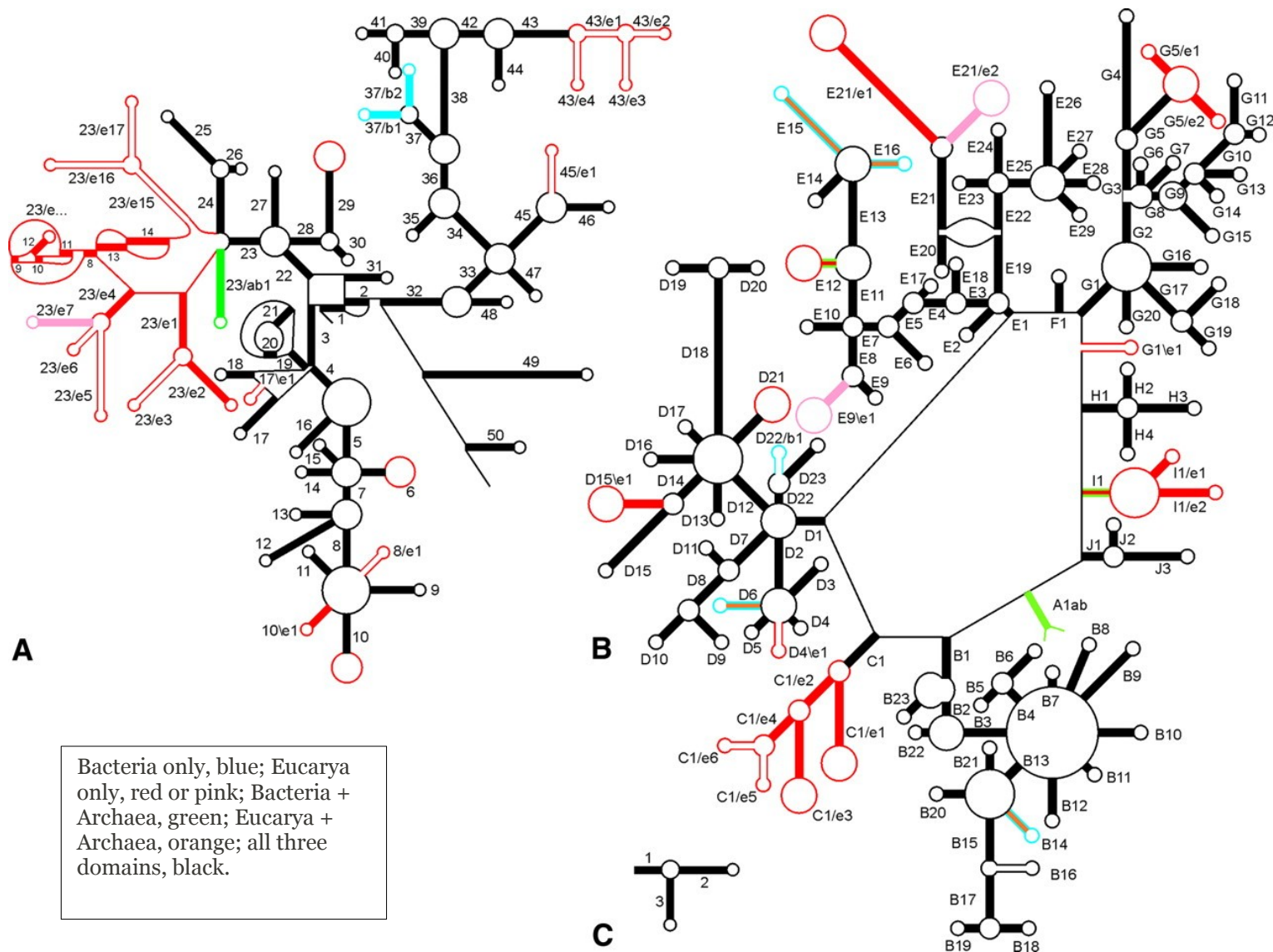


# Potential problem with ribosomal RNA? May be too conserved

- Ribosomal RNA is very conserved even between Bacteria and Eukaryots.
  - Is there a region in Ribosomal RNA that can be used
  - Yes: V4 and V9 regions are relatively commonly present in Eukaryots and absent in Bacteria.



**Figure 1. Secondary structure and helix numbering of SSU rRNA (A), LSU rRNA (B) and 5S rRNA (C).**

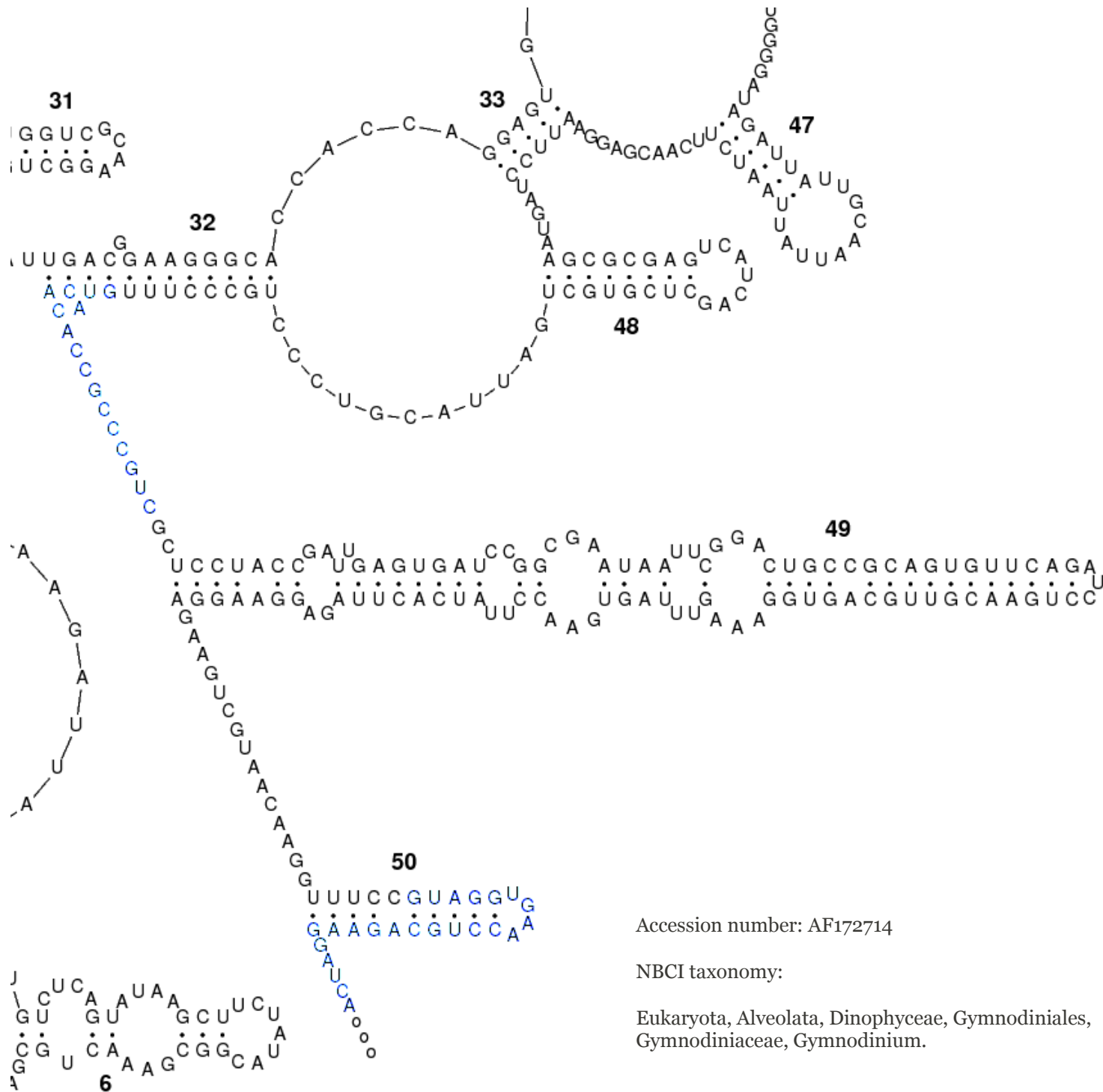


Wuyts J et al. Nucl. Acids Res. 2001;29:5017-5028





# Gymnodinium breve SSU rRNA secondary structure model



Accession number: AF172714

NBCI taxonomy:

Eukaryota, Alveolata, Dinophyceae, Gymnodiniales,  
Gymnodiniaceae, Gymnodinium.

>ENA|AF172714|AF172714.1 *Gymnodinium breve* strain CCMP718  
18S small subunit ribosomal RNA gene, partial sequence.  
aacctggttgatcctgccagtagtcatatgcttgtctcaaagattaagccatgcatgtct  
cagtataagcttctatacggcgaaactgcgaatggctcattaaaacagttatagtttatt  
tgatggtcattcattacatggataactttggaaattctagagctaatacatgcccctgctg  
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aacgaacgagaccttaacctgctaaatagttacacgtaacttcggttacgtgggcaactt  
cttagagggactttgctgttctaacgcaaggaagtttgaggcaatagcaggtctgtgatg  
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tgcccggaaggttgggtaatctttttaaacgcatcgtgatggggatagattattgcaat  
tattaatcttcaacgaggaattcctagtaagcgcgagtcacagctcgtgctgattacgt  
ccctgcccttt**gtacacaccgcccgtc**gctcctaccgattgagtgatccggcgaataatt  
cggactgccgagtggtcagatcctgaacgttgagtgaaagtttagtgaaaccttatca  
cttagaggaaggagaagtcgtaacaaggtttcc**gtaggtgaacctgcagaaggatca**

Red: v4 region  
Avg. 382 nt

Blue: v9 region  
Avg. 126 nt

# Some statistics on Silva



<http://www.arb-silva.de/>

V4: CCAGCA[GC]C[CT]GCGGTAATTCC    T[CT][AG]ATCAAGAACGAAAGT

Up to 4 mismatches per primer:

DB 262092 seq. // 219921 Bacteria // 10302 Archae // 31809 Eukaryota

->> 29303 seq. // 944 Bacteria // 16 Archae // 28296 Eukaryota

V9: GTACACACCGCCCGTC    GTAGGTGAACCTGCAGAAGGATCA

->> 11632 seq. // 5447 Bacteria // 144 Archae // 6025 Eukaryota

# Number of mismatches on SSU RNA

## V9

Bacteria							Eukaryota							
	15	16	20	21	22	23		15	16	20	21	22	23	24
12	0	0	6	1	0	0	12	0	0	0	1	3	15	3
13	0	0	6	1	0	0	13	0	0	2	5	2	7	3
14	0	0	10	0	0	0	14	0	0	1	10	5	7	3
15	6	0	227	42	1	0	15	6	0	6	3	17	70	49
16	0	46	5054	35	0	2	16	0	159	295	156	370	2729	2084

## V4

Bacteria				Eukaryota					
	14	15	18		14	15	16	17	18
14	3	0	0	14	0	0	0	0	2
16	17	0	0	15	0	2	0	0	3
17	67	7	0	16	3	1	13	13	49
18	835	9	2	17	0	8	8	35	99
19	1	0	0	18	10	33	26	51	333
20	0	0	3	19	211	416	142	163	870
				20	106	486	790	4918	19373



# Available Barcode Data

- V9 loop, DNA
- 6 stations
- 2 depths
  - Subsurface
  - DCM
- 4 size fractions:
  - 0.8-5  $\mu\text{m}$
  - 5-20  $\mu\text{m}$
  - 20-180  $\mu\text{m}$
  - 180-2000  $\mu\text{m}$









## 1. Find primers

GTACACACCGCCGGTCGTTCTTACCGATTGAATGATCCGGTGAGCCGCCCGatggagtcactgogtccggttttcccgacaatgagaccttccgtgaagcttcgtaccctaaaccttta  
tctatcttagaggaaggagaagtcgtaacaaggtccttccccgtaggtgaacctgcagaagg

## 2. Remove primers

GTTCTTACCGATTGAATGATCCGGTGAGCCGCCCGatggagtcactgogtccggttttcccgacaatgagaccttccgtgaagcttcgtaccctaaacctttatctatcttagaggaag  
gagaagtcgtaacaaggtccttcccc

## 3. Extract corresponding qualities

37 39 39 37 37 39 39 37 35 35 37 37 37 37 37 39 37 37 37 37 37 37 37 37 37 37 32 25 25 25 25 28 32 33 32 32 32 28 28 19  
19 19 23 24 18 17 17 23 16 16 11 14 14 14 12 12 14 11 14 19 19 17 19 25 19 17 19 26 27 17 17 13 13 13 13 21 21 16 16 18  
13 16 20 22 22 13 12 12 12 12 12 11 11 13 16 17 12 12 12 18 12 12 12 18 22 28 20 16 11 12 11 11 11 11 11 11 11 11 16  
17 28 20 18 11 11 11 17 13 12 12 12 20 20 22 22 20 16 16 17 18 19 19 18

## 4. Convert quality values to Expected number of errors.

$$EE = \sum_i 10^{-Q_i/10}$$

## 5. Dereplicate sequences

>NPGJ8MOYR06G3N03\_724 EE=0.015525 MD5=4e38e8ced9070952b314e1880bede1ca L=127 S=T0099  
GCTACTACCGATTGAACGTTTTAGTGAGGTCTTCGGACTGTTTGGTAGTCGGATCACTCTGACTGCCTGGCGGGAAGACG  
ACCAAACGTAGCGTTTTAGAGGAAGTAAAAGTCGTAACAAGGTTTTCC

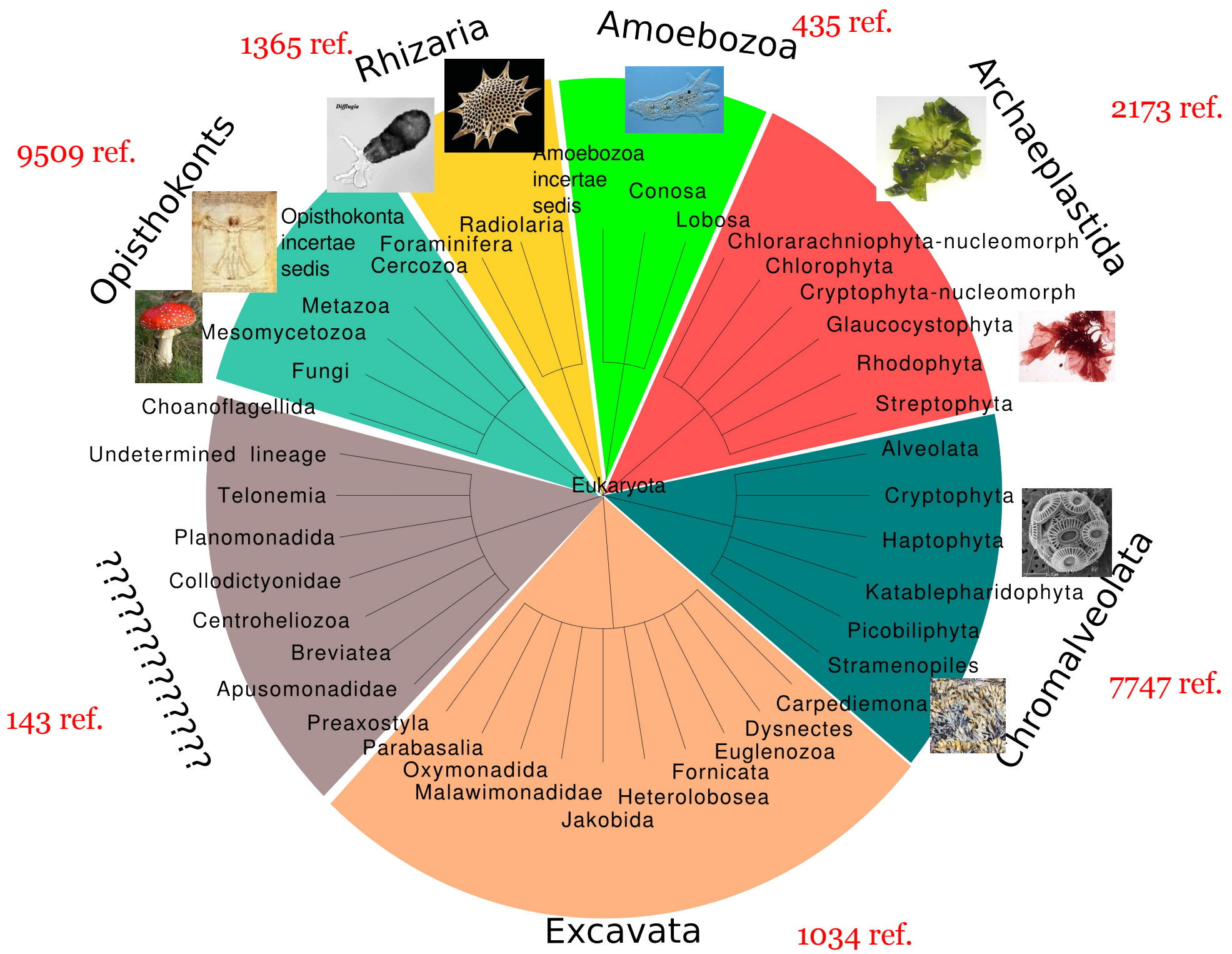
## 5. Filter sequences:

Sequences which are unique (within or between samples), with  $EE/L > 1\%$  (more than 1 % of error) are disregarded.

In the above case  $EE = 3.93$ , sequence is disregarded.

# Sequence assignation: giving a name to those sequences

- Sequence assignation is performed from a dedicated Reference database:
  - Built from a curated database: we use Laure Guillou's 18S database, soon to be incorporated in Silva.
  - We extract (from primer search with degeneracy), parts of the sequence corresponding to the targeted fragment.
    - One primer set = one reference database
    - guided search: less false positives, faster processing



# Example reference database

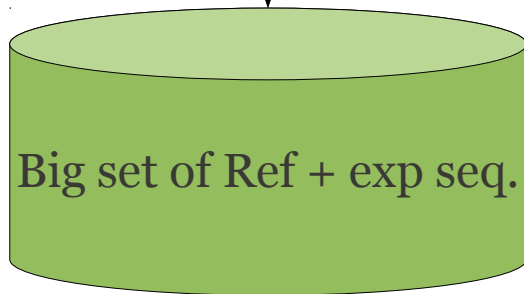
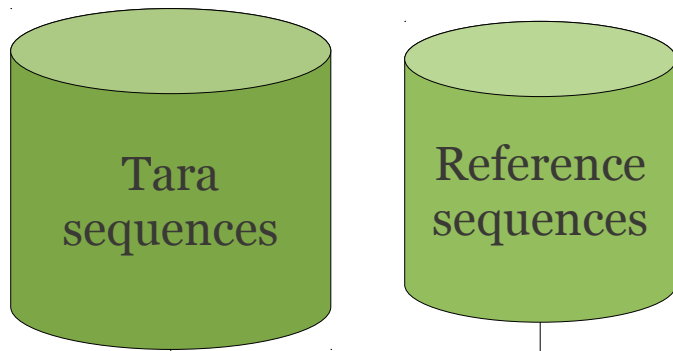
Identifier

Sequence

Lineage

X53904 |  
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ttaccgtaggtgaacctgcggaaggatcattg | Eukaryota | Archaeplastida | Chlorophyta | Chlorophyceae | Chlorophyceae | CW\_Chlamydomonadales |  
Volvox | carteri | \*  
X73991 |  
gctcctaccgattggatgtgctgggtgaagcgttcggattggagtcactcggcggtcgtccgccggtggcaccgagaagttcgttaaaccctcccacctagaggaaggagaagtcgtaa  
caaggtctcc | Eukaryota | Archaeplastida | Chlorophyta | Trebouxiophyceae | Trebouxiophyceae | Watanabea Clade | Watanabea | reniformis | \*  
U13985 |  
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tttccgtagtgaacc | Eukaryota | Archaeplastida | Chlorophyta | Chlorophyceae | Chlorophyceae | CW\_Chlamydomonadales | Chlamydomonas |  
dysosmos | \*  
X62441 |  
gctcctaccgattgggtgtgctgggtgaagtgttcggattggcgaccggggcggtctccgctctcggccgcccgagaagttcattaaaccctcccacctagaggaaggagaagtcgtaaca  
aggtttcc | Eukaryota | Archaeplastida | Chlorophyta | Trebouxiophyceae | Trebouxiophyceae | Chlorellales | Chlorella | sorokiniana | \*  
U70784 | gctcctaccgattgggtgtgctgggtgaagtgttcggattggcttcaggtgatggcaacatcgcttggctgagaagttcattaaaccctcccacctagaggaagg |  
Eukaryota | Archaeplastida | Chlorophyta | Chlorophyceae | Chlorophyceae | CW\_Chlamydomonadales | Lobo-chlamys | segnis | \*  
AJ132619 |  
gctcctaccgattggattggctgggtgaagcgttcggattgtgggttcggagcgggttcgccgcttctgggcttgcgagaagttcgttaaaccctcctatctagaggaaggagaagtcgtaac  
aaggtttccgt | Eukaryota | Archaeplastida | Chlorophyta | Prasinophyceae | Prasi-no-Clade V | Pycnococccaceae | Pseudoscourfieldia |  
marina | \*  
AB017123 |  
gctcctaccgattgaaatgggtccgggtgaaatgttcggaccgcccggagacggacgggttcgctgtcagcctcgctgcccgggaagttcattaaacccttatcatttagaggaaggagaagtcgta  
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X73993 |  
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aggtttcc | Eukaryota | Archaeplastida | Chlorophyta | Trebouxiophyceae | Trebouxiophyceae | Chlorellales | Chlorella | sorokiniana | \*  
U13984 |  
gctcctaccgattgggtgtgctgggtgaagtgttcggattgacttcagcgggggcaactctgctggttggagaagatcattaaaccctcccacctagaggaaggagaagtcgtaacaagg  
tttccgtagtgaacc | Eukaryota | Archaeplastida | Chlorophyta | Chlorophyceae | Chlorophyceae | CW\_Chlamydomonadales | Chlamydomonas |  
applanata | \*  
X74001 |  
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aggtttcc | Eukaryota | Archaeplastida | Chlorophyta | Trebouxiophyceae | Trebouxiophyceae | Chlorellales | Chlorella | sorokiniana | \*





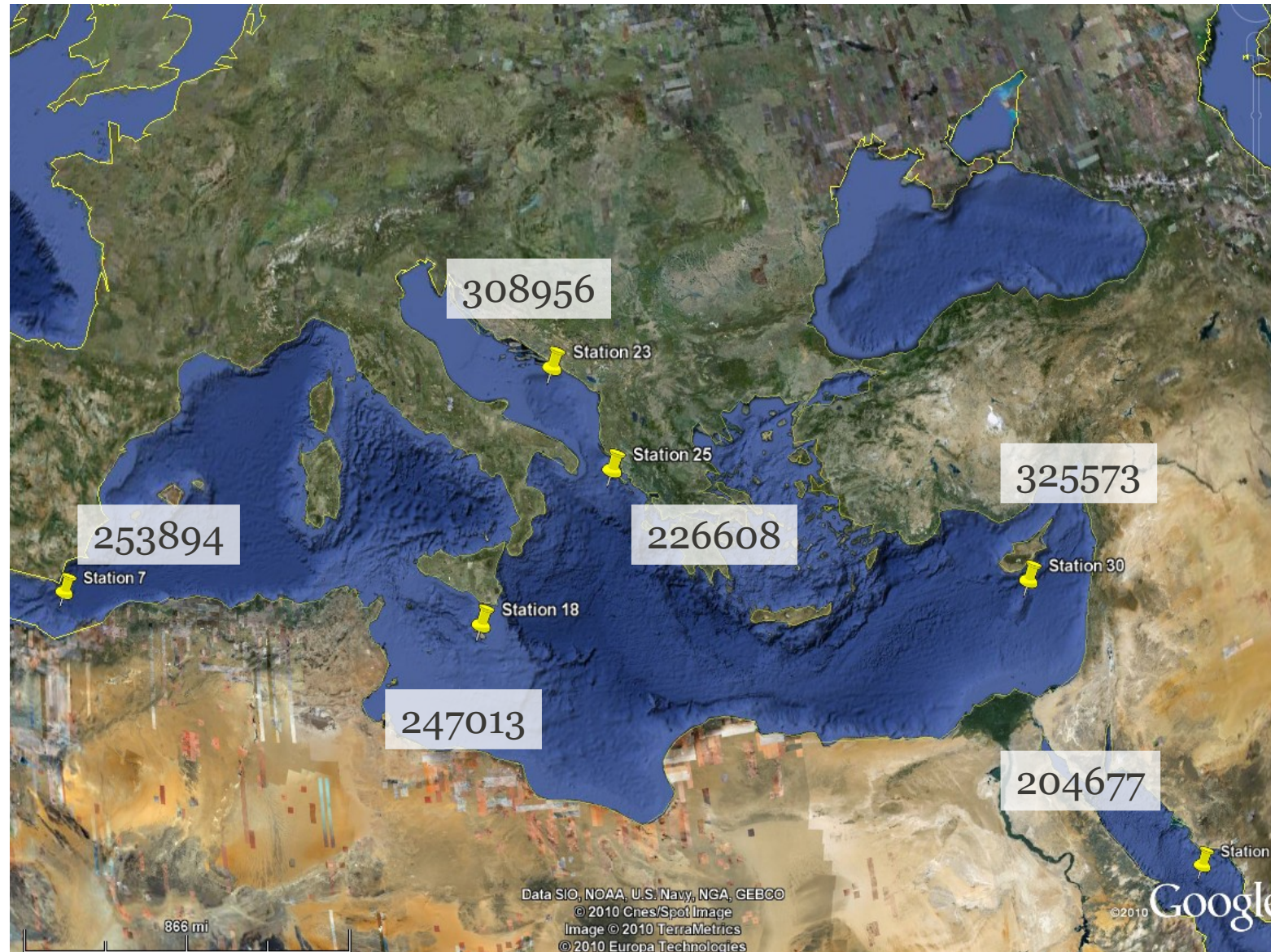
Run uclust at several clustering level: 85% to 100%

Seqname	abund.	sample	c85	c86	c87	..	c100
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AY664978	0	REF	5	1289	1423	..	239059
NPGJ8MOYR03DPXD1_1	1	T0081	5	1289	1423	..	250630
NPGJVVFSSH02CB4FG_28	28	T0028	5	1289	1423	..	258867
NPGJA7C3N03GGBKZ_5	5	T0012	5	1289	1423	..	275350
NPGJ8MOYR02B3DC9_78	78	T0077	5	1289	1423	..	165798

Perform assignation for each sequence by looking at level c100 if there is a Reference sequence in the same cluster. If not look at level c99, and so on.

# Preliminary analysis of the Barcode datasets

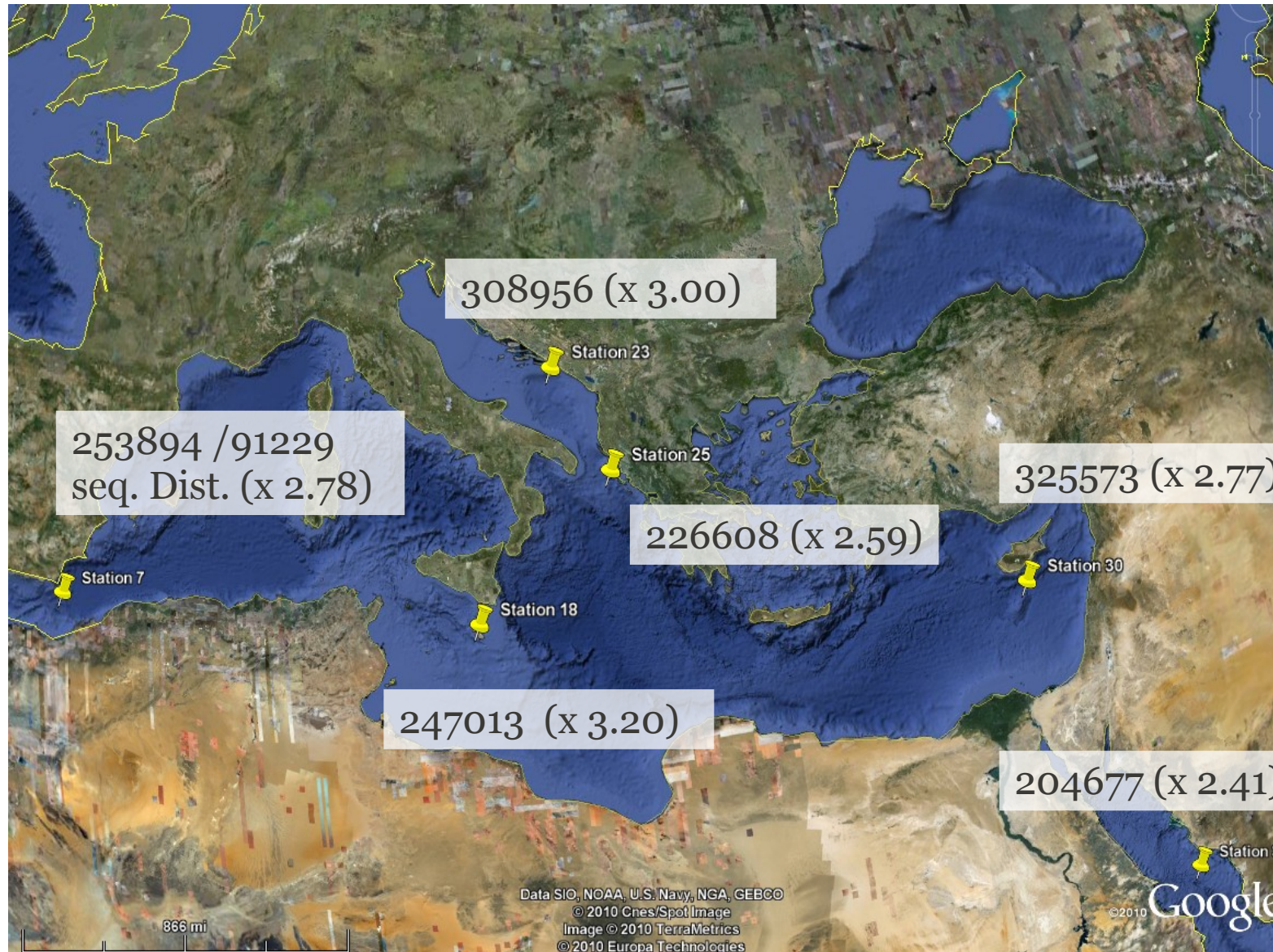
## Number of valid sequences





# Preliminary analysis of the Barcode datasets

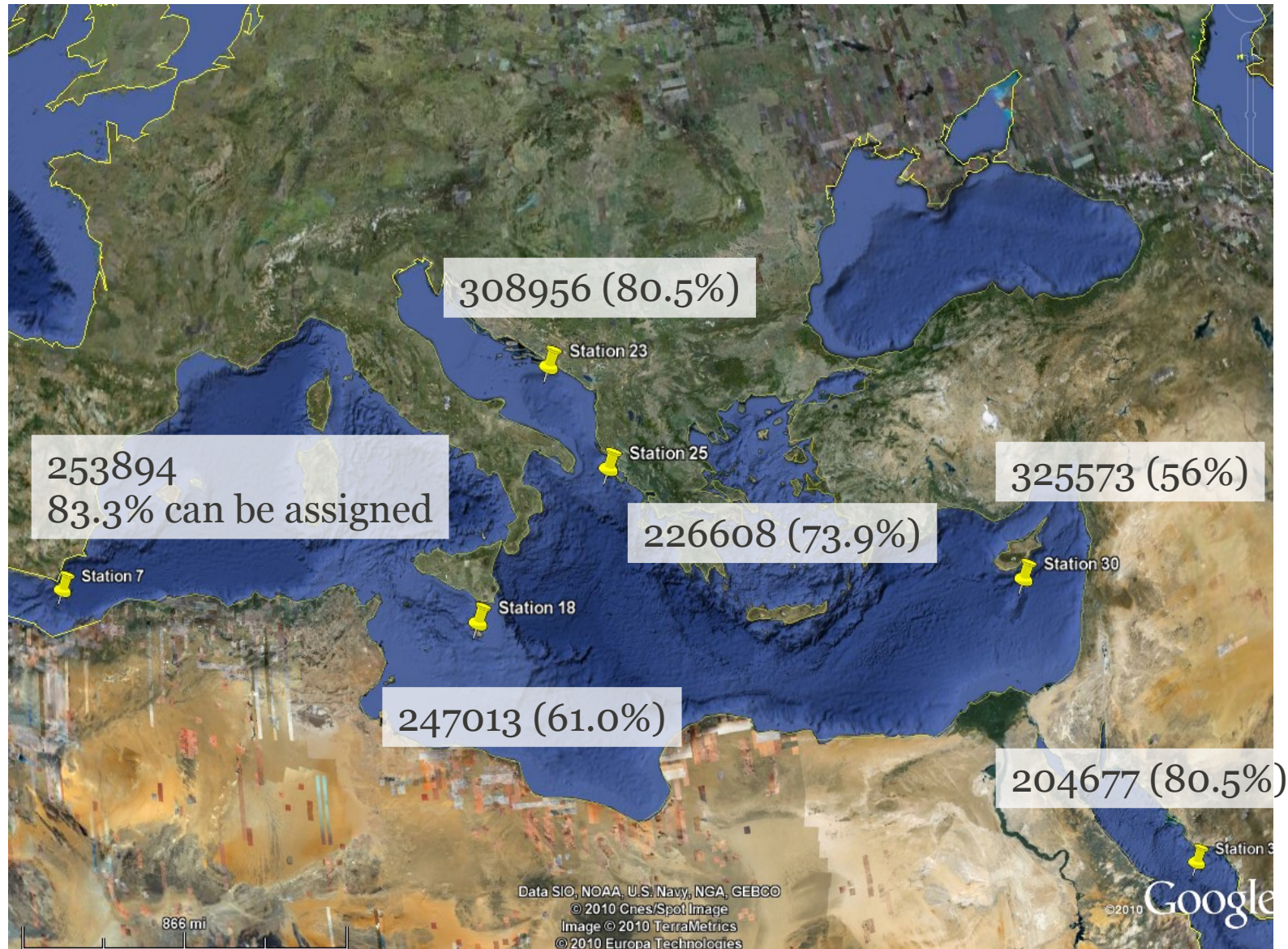
## Multiplicity of sequences



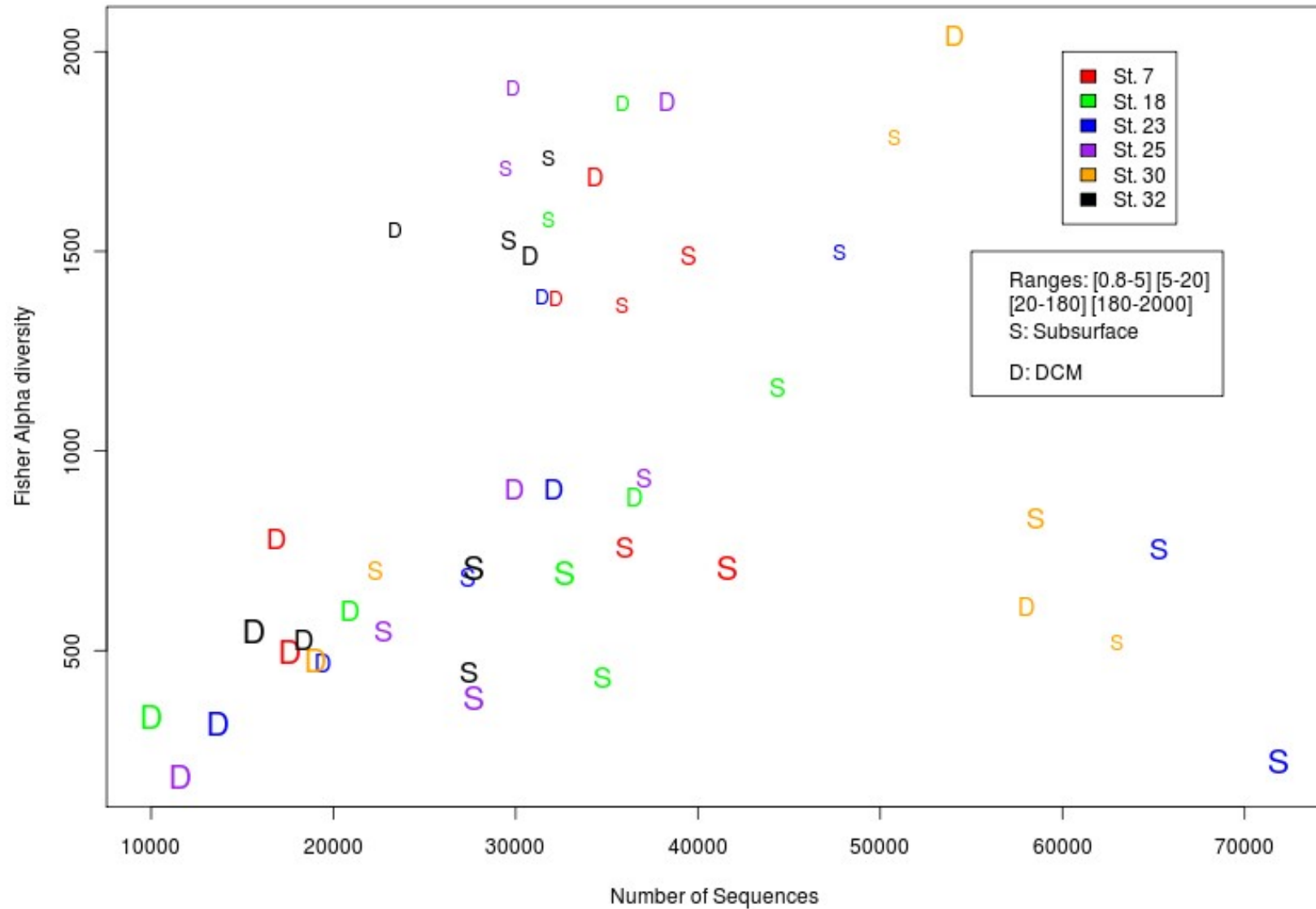


# Preliminary analysis of the Barcode datasets

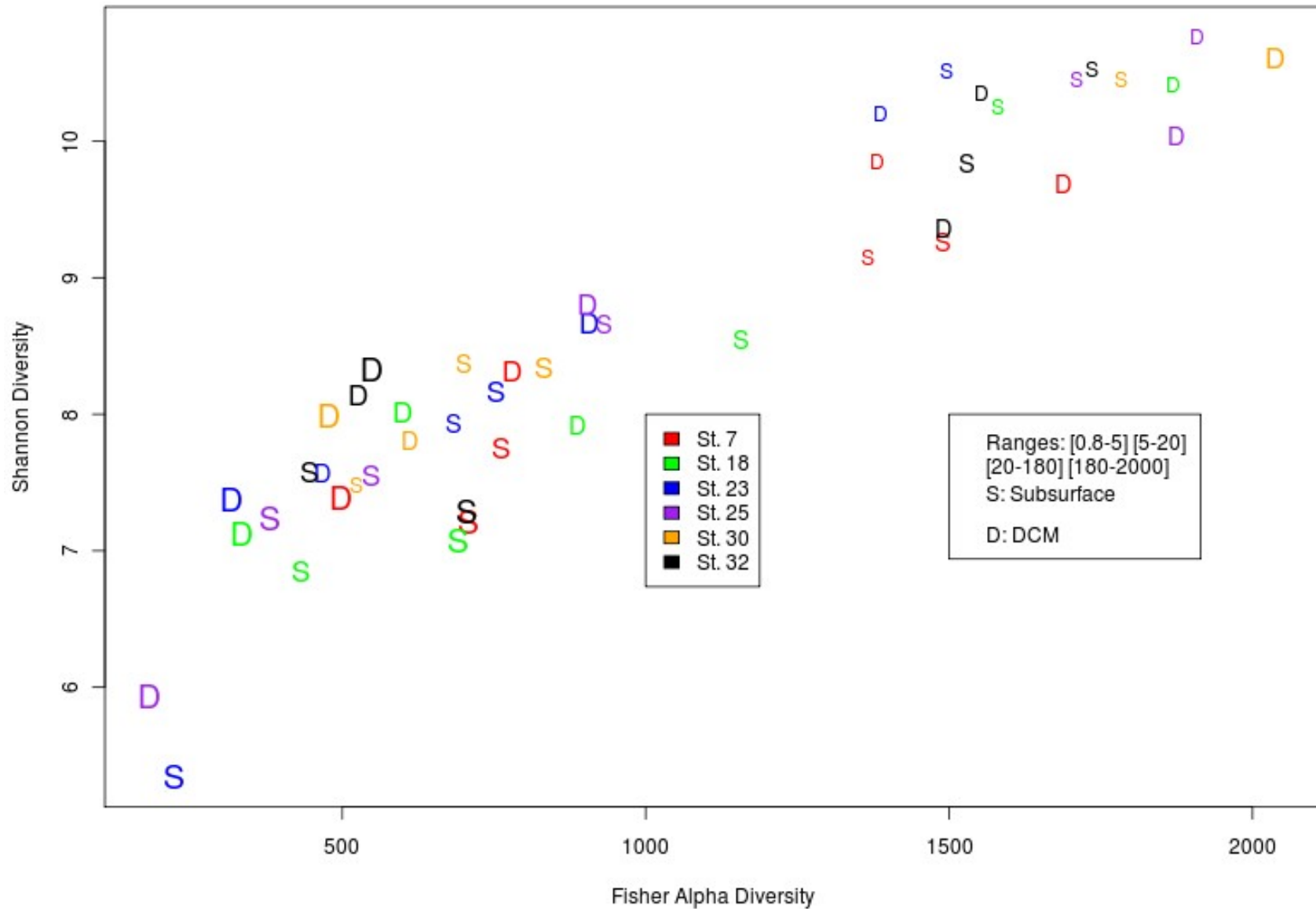
## Fraction of assigned sequences



# Small size fractions are more diverse



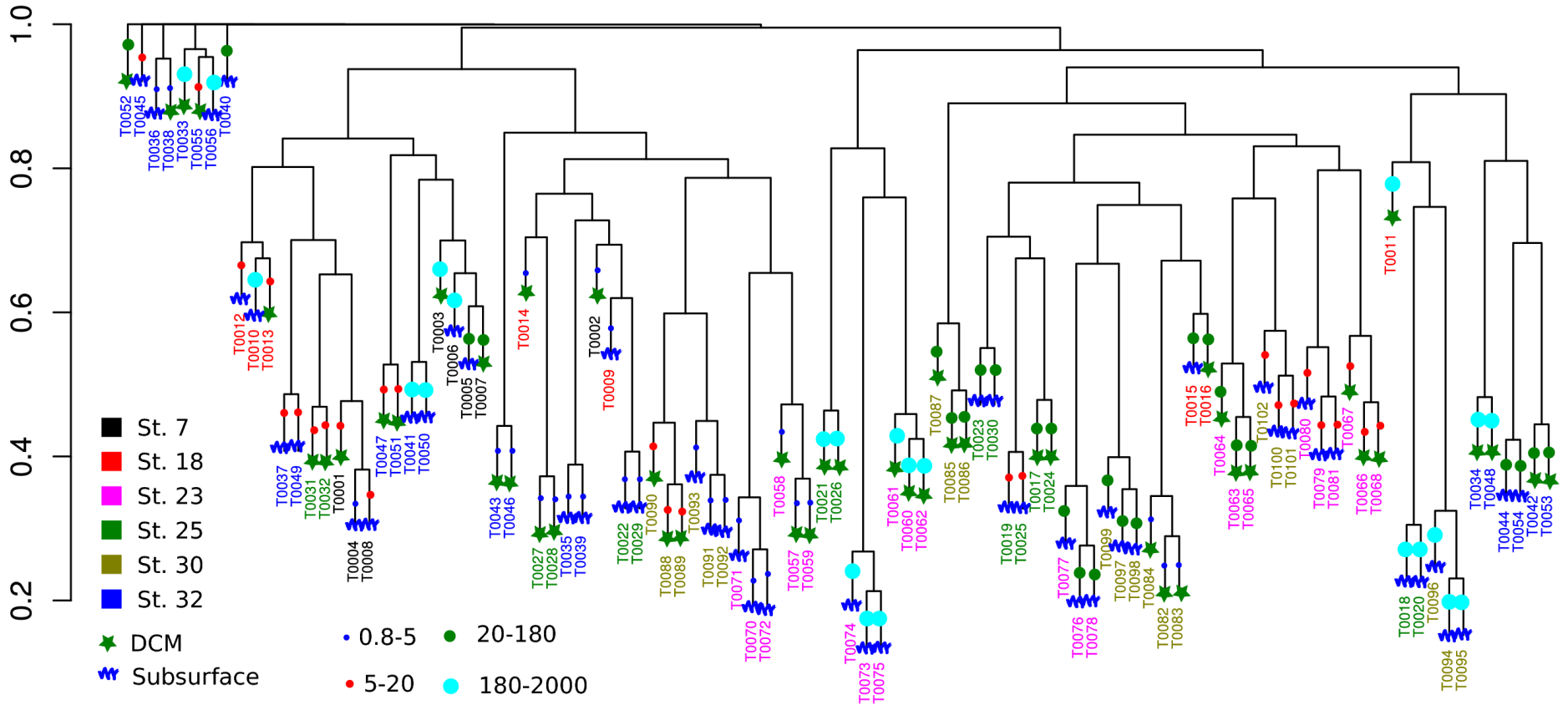
# Small size fractions are more diverse





# Clustering of samples

Jaccard distance , cluster at 97%



Distance between sample is calculated for sequence clusters at 97% identity (uclust). We used Jaccard distance:  $J(A,B) = 1 - \frac{|A \cap B|}{|A \cup B|}$ .

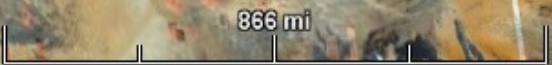
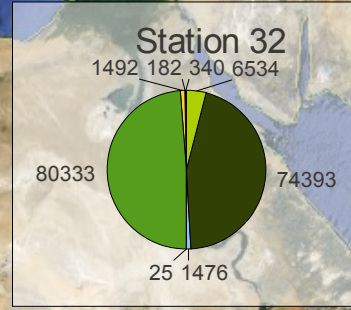
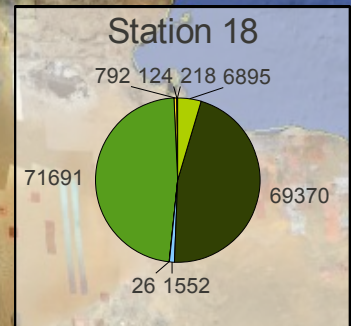
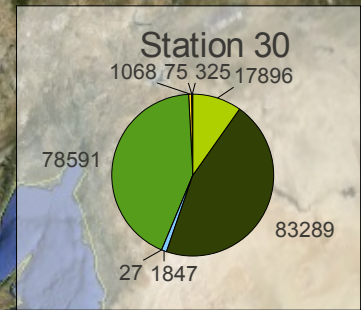
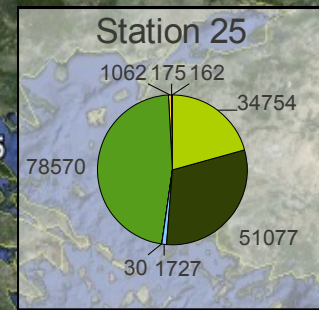
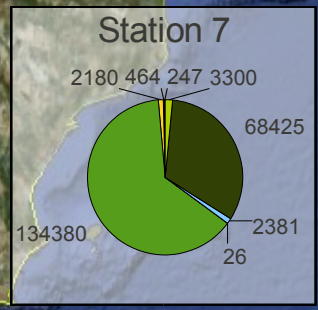
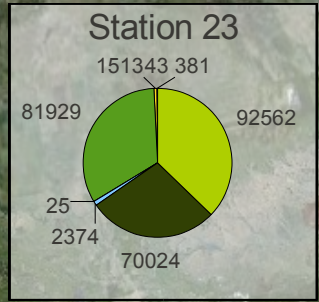
3 sets of seq. data for St 32 (1 run of sequencing failed-> cluster on the top left)

Small size fraction of all stations cluster together (one exception, Too4?)



# Assignations at the supergroup level

- \*
- Amoebozoa
- Archaeplastida
- Chromalveolata
- Excavata
- "Incertae sedis Eukaryota"
- Opisthokonta
- Rhizaria



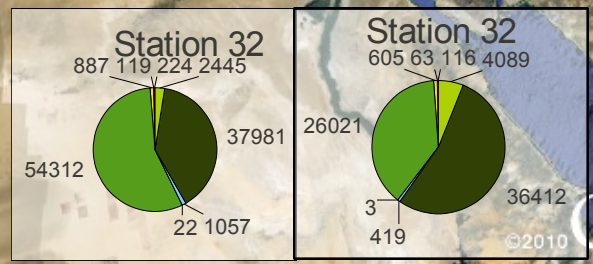
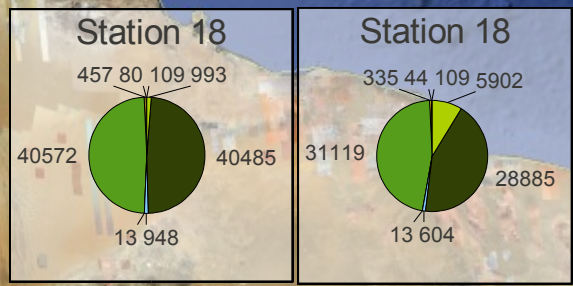
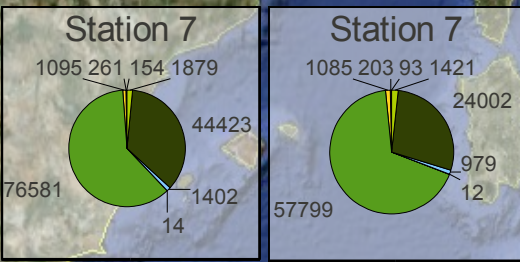
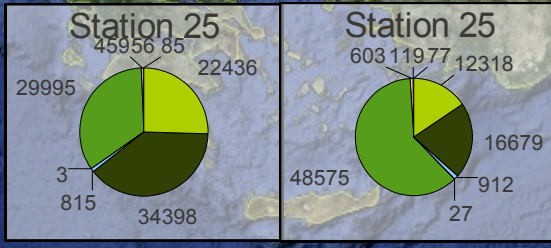
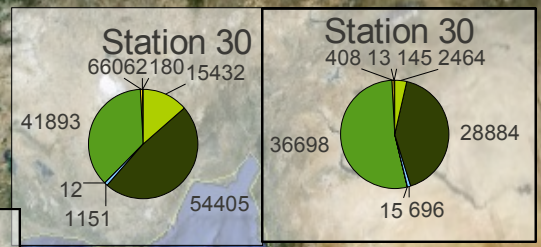
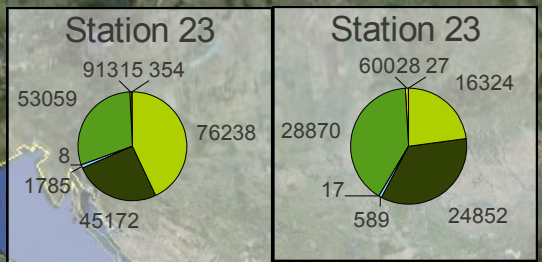
Data SIO, NOAA, U.S. Navy, NGA, GEBCO  
 © 2010 Cnes/Spot Image  
 Image © 2010 TerraMetrics  
 © 2010 Europa Technologies

©2010 Google



# Assignations at the supergroup level, Subsurface (left) vs. DCM (right)

- \*
- Amoebozoa
- Archaeplastida
- Chromalveolata
- Excavata
- "Incertae sedis Eukaryota"
- Opisthokonta
- Rhizaria



866 mi



# Interactive Web Interface



Choose Samples

Choose Taxonomy

Configure

[Help](#)

## Sample selector

Sequences will be selected according to the sample selection *AND* the taxonomy selection *AND* the configuration choice you have made. Pending a better sample selection interface, a more detailed description of the samples is available ([here](#))

Sample:

- T0051  T0052  T0053  T0054  T0055  T0056  T0057  T0058  T0059  T0060
- T0061  T0062  T0063  T0064  T0065  T0066  T0067  T0068  T0070  T0071
- T0072  T0073  T0074  T0075  T0076  T0077  T0078  T0079  T0080  T0081
- T0082  T0083  T0084  T0085  T0086  T0087  T0088  T0089  T0090  T0091
- T0092  T0093  T0094  T0095  T0096  T0097  T0098  T0099  T0100  T0101



Primer:  Sampling Session:  Molecule type:

Depth:  Size Fraction:

[Click to see \(expire in 5 minutes\)](#)

[Click to see Pie Chart \(expire in 5 minutes\)](#)

Choose Samples

**Choose Taxonomy**

Configure

[Help](#)

## Taxonomy selector

Choose here the species you want to retrieve. Each selected level will update the menu for the next level. Sequences will be selected according to this taxonomy selection *AND* the sample selection *AND* and your configuration choice.

If "all" is selected at all levels, even sequences that could not be assigned will be returned. For each taxonomic level, the menu items have the following format "Taxon name (number of sequences / number of distinct sequences)"

Taxonomy level

- \* (354/144)
- Amoebozoa (15/12)
- Archaeplastida (913/521)
- Chromalveolata (53059/23116)
- Excavata (8/8)
- Incertae sedis Eukaryota (1785/742)
- Opisthokonta (45172/12367)
- Rhizaria (76238/14621)**
- all

Get Selected Sequences

Include Ref. Sequences

Get Sequence by ID

Show species distribution

[Click to see \(expire in 5 minutes\)](#)

Show Seq. Distribution per Sample

Pie Chart

[Click to see Pie Chart \(expire in 5 minutes\)](#)

[Choose Samples](#)[Choose Taxonomy](#)[Configure](#)[Help](#)

## Taxonomy selector

Choose here the species you want to retrieve. Each selected level will update the menu for the next level. Sequences will be selected according to this taxonomy selection *AND* the sample selection *AND* and your configuration choice.

If "all" is selected at all levels, even sequences that could not be assigned will be returned. For each taxonomic level, the menu items have the following format "Taxon name (number of sequences / number of distinct sequences)"

▾
  ▾
  ▾
  ▾

▾
  ▾
  ▾
  ▾

\* (29/19)

 ▾



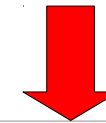
[Click to see \(expire in 5 minutes\)](#)

 ▾
[Click to see Pie Chart \(expire in 5 minutes\)](#)

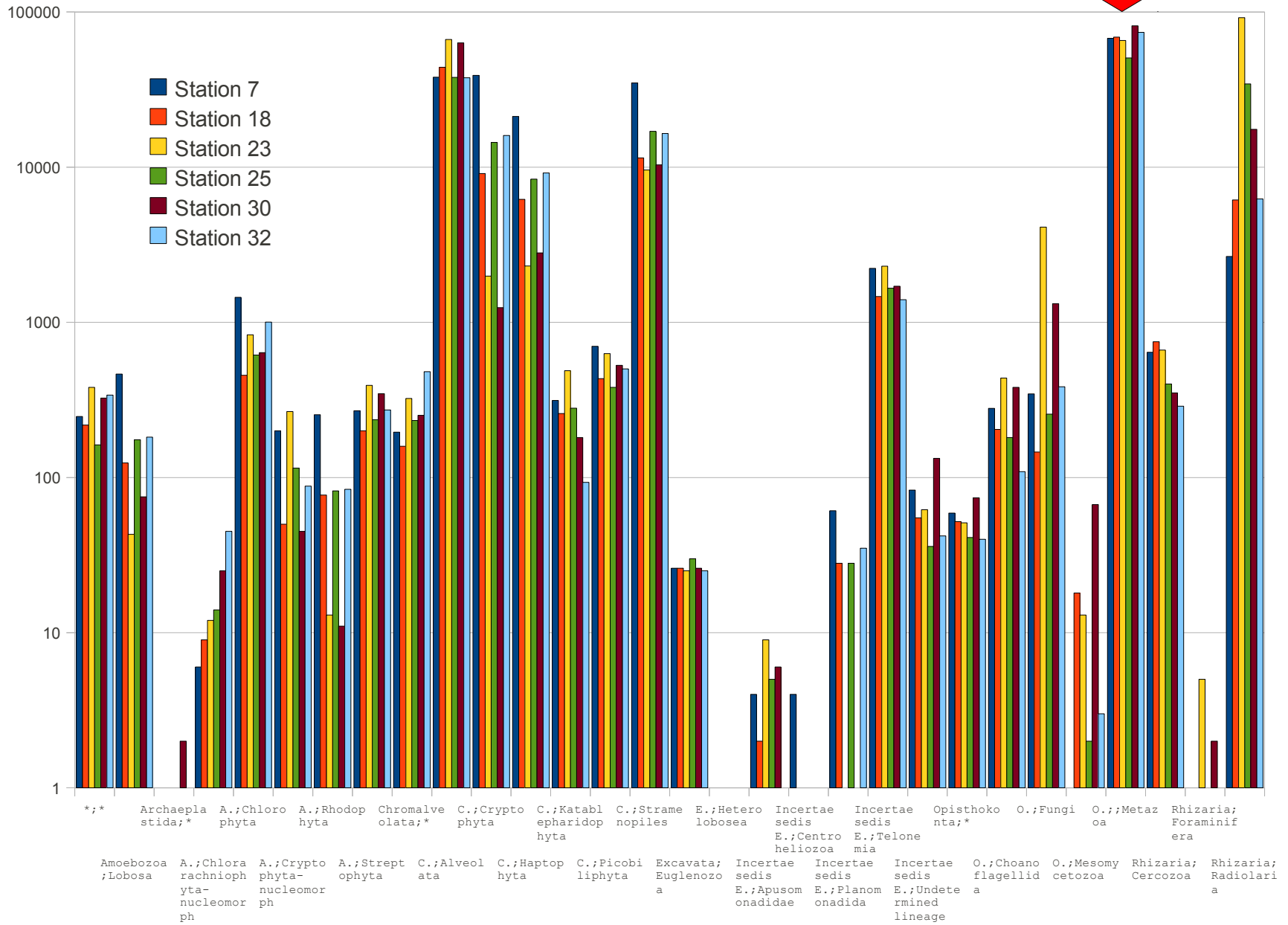
[Click to see \(expire in 5 minutes\)](#)



# Repartition of assigned sequences per division

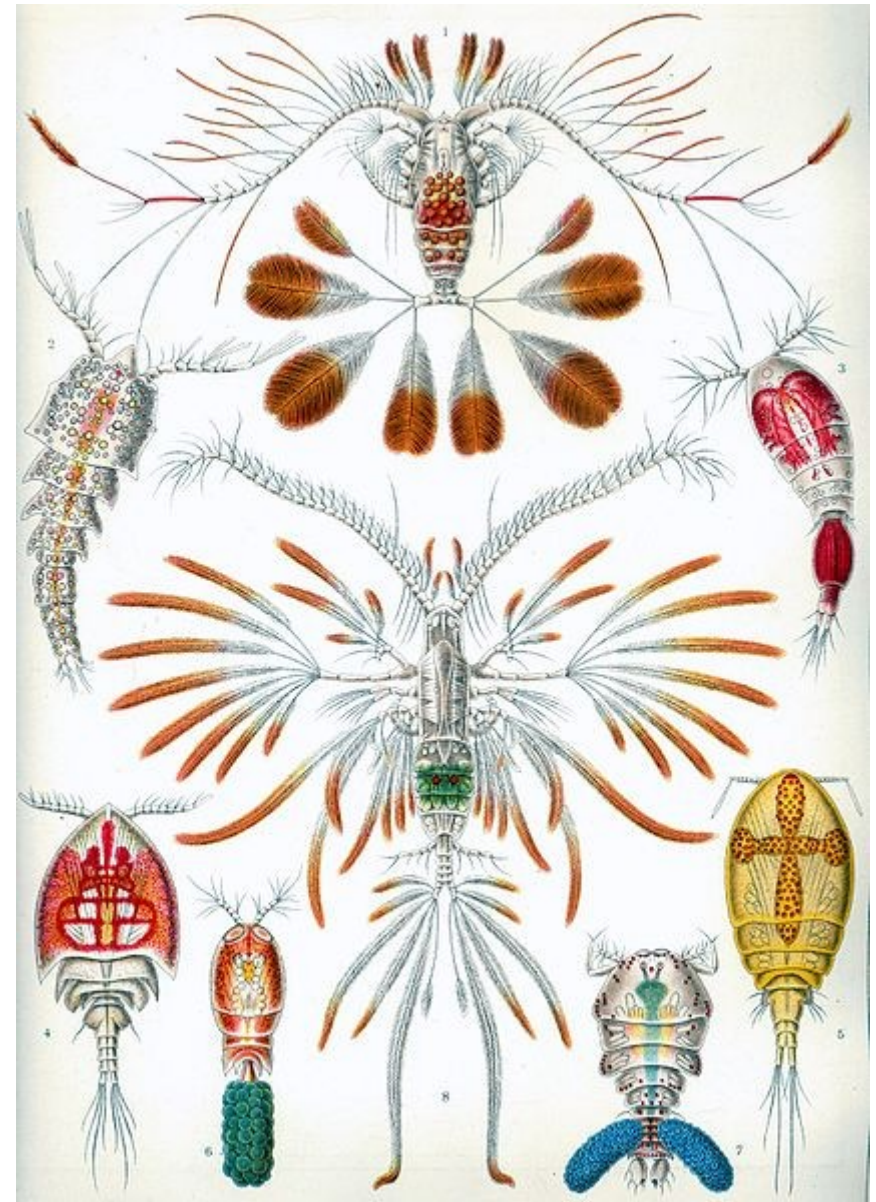


Metazoa



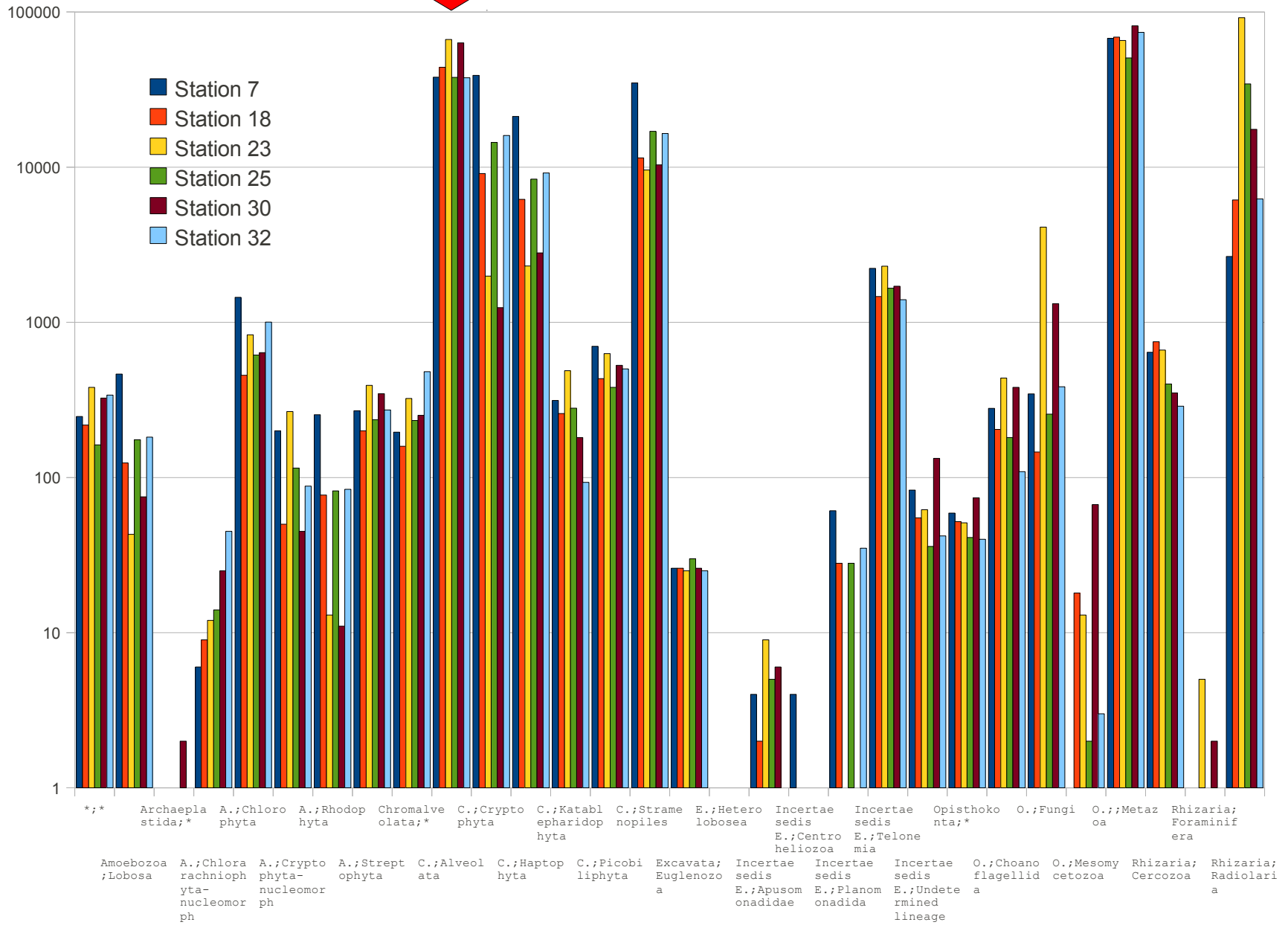
Kunstformen der Natur (1900), Ernst Haeckel

- 294714 Arthropoda Crustacea
- 54281 Cnidaria Cnidaria
- 15201 Urochordata Urochordata
- 11263 \* \*
- 10044 Annelida Annelida
- 8888 Chaetognatha Aphragmophora
- 4148 Ctenophora Ctenophora
- 3522 Mollusca Gastropoda
- 1452 Platyhelminthes Turbellaria
- 673 Craniata Craniata

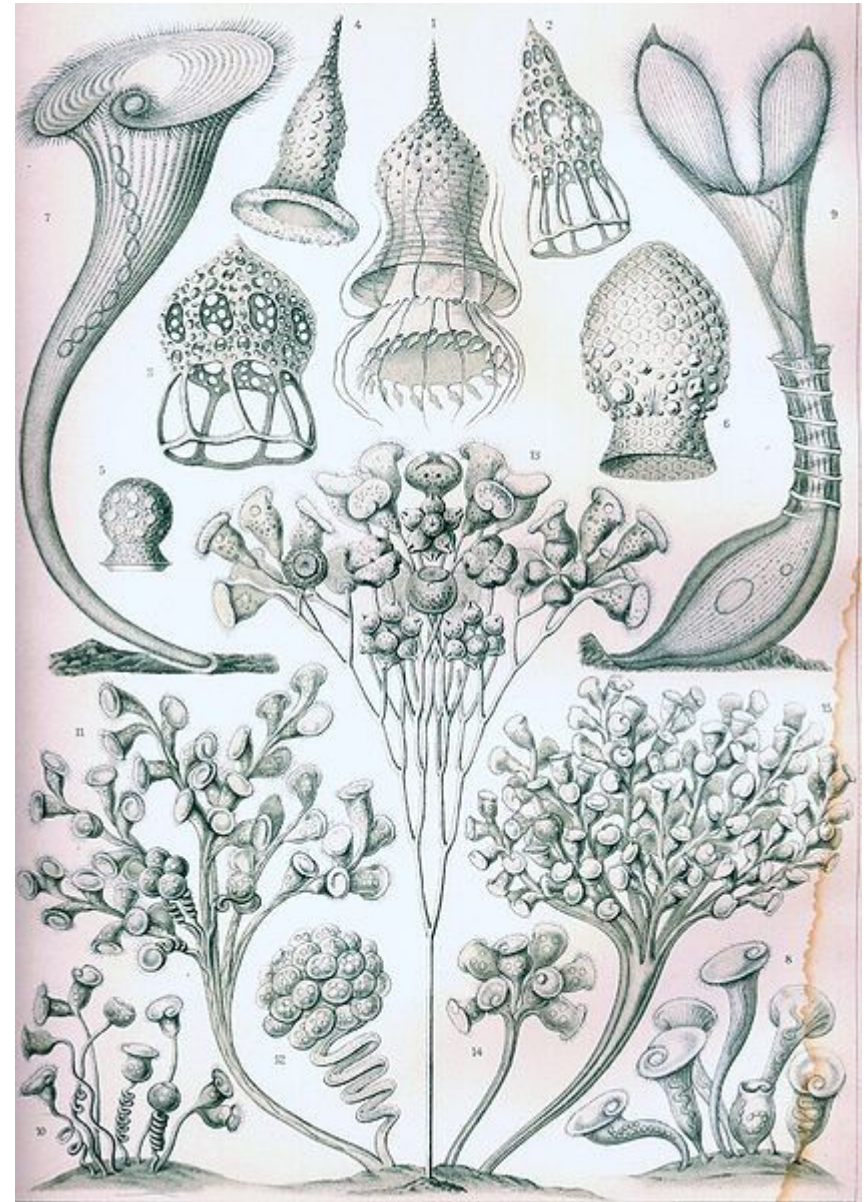


# Alveolata

Replication of assigned sequences per division



140728 Dinophyceae Dinophyceae  
111073 Dinophyceae Syndiniales  
12998 Dinophyceae \*  
8937 Ciliophora Oligotrichea  
6916 \* \*  
1453 Perkinsea Perkinsida  
1386 Ciliophora Oligohymenophorea  
929 Apicomplexa Coccidia  
564 Ciliophora Colpodea and Nassophorea  
519 Apicomplexa Cryptosporidium

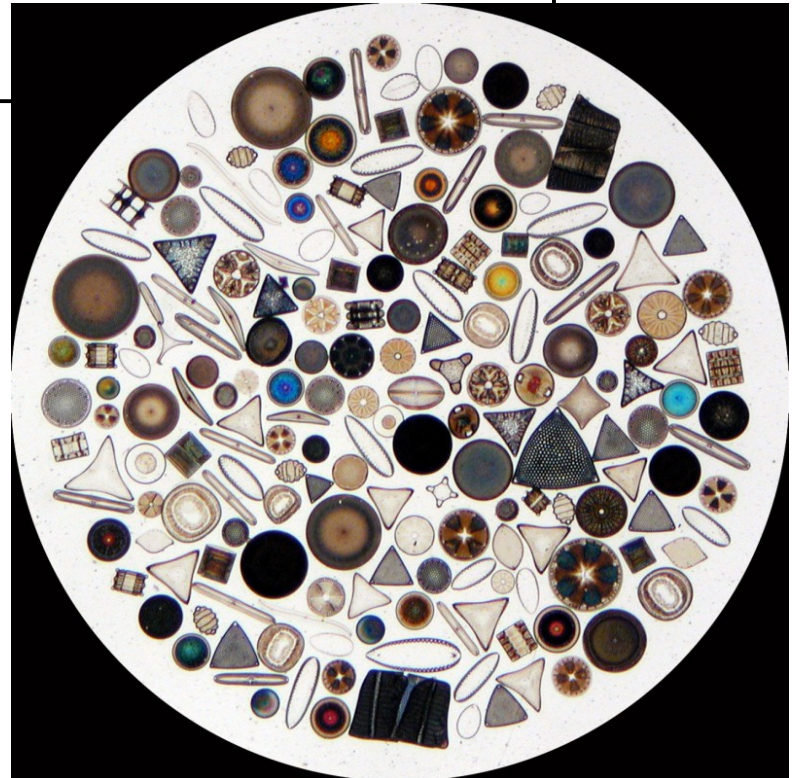
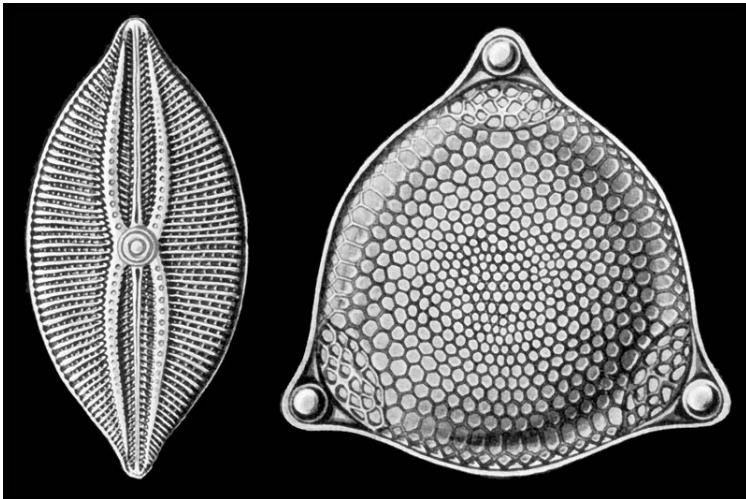




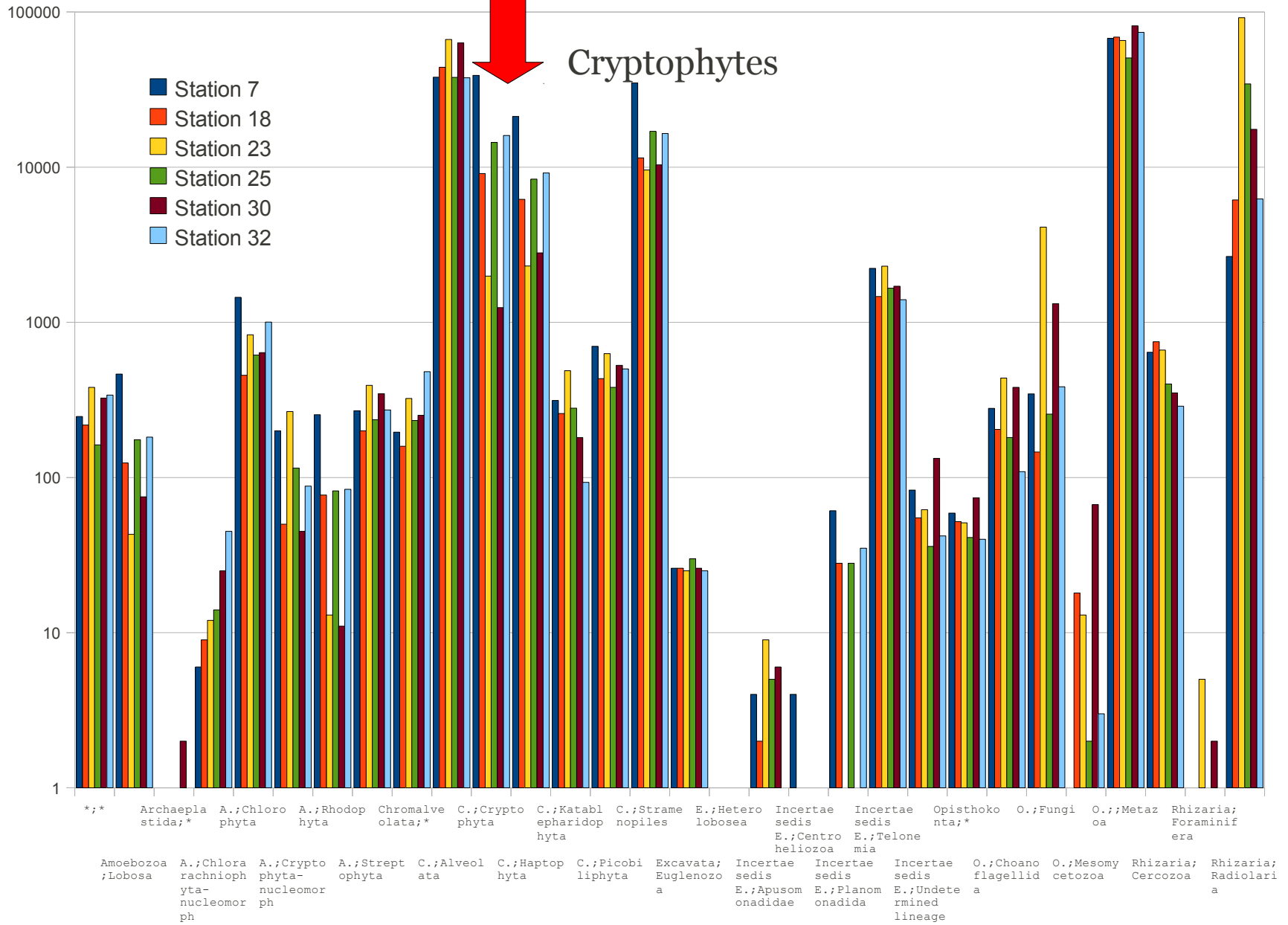


32264 Bacillariophyta Centric diatoms\_Coscinodiscophyceae  
16746 Pinguiphyceae Pinguiphyceae  
9992 MAST-1 MAST-1  
9934 Bacillariophyta Pennate diatoms  
6232 MAST-3 MAST-3  
4662 Chrysophyceae and Synurophyceae Chrysophyceae and Synurophyceae  
4605 MAST-4 MAST-4  
4229 \* \*  
2545 Pelagophyceae Pelagophyceae  
1295 MAST-9 MAST-9

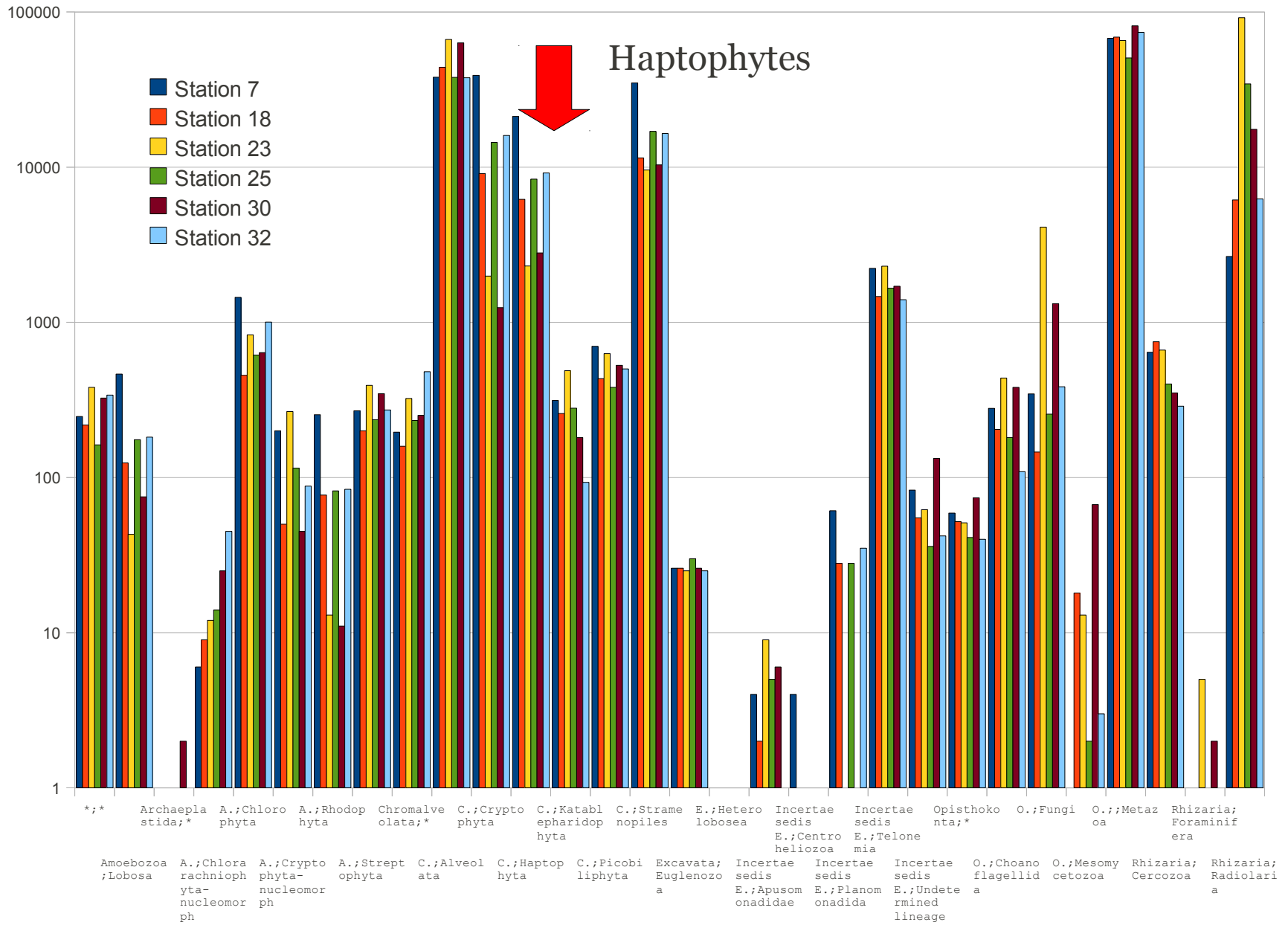
Kunstformen der Natur (1900), Ernst Haeckel



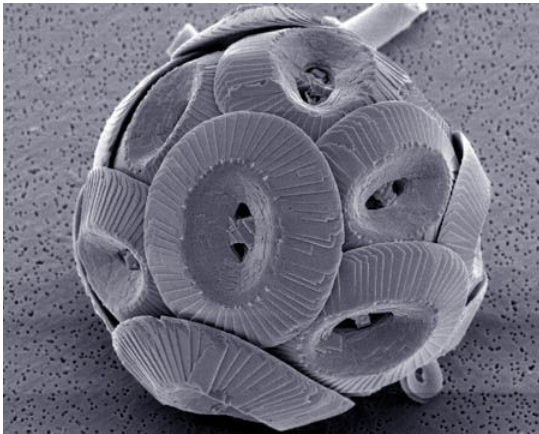
Repartition of assigned sequences per division



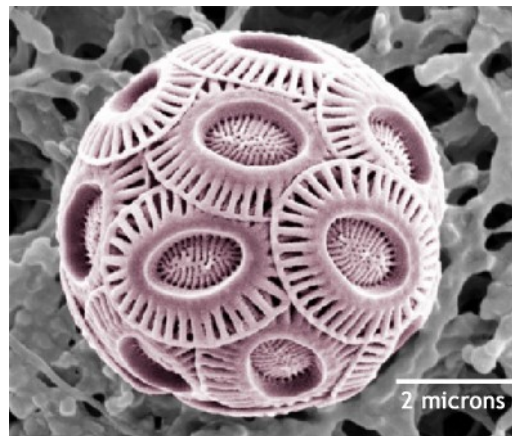
# Repartition of assigned sequences per division



30052	Prymnesiophyceae Prymnesiophyceae *
12775	Pavlovophyceae Pavlovophyceae *
4253	Prymnesiophyceae Prymnesiophyceae Chrysochromulina group
1147	Prymnesiophyceae Prymnesiophyceae Pleurochrysis group
840	Prymnesiophyceae Prymnesiophyceae Emiliana and Gephyrocapsa group
245	Prymnesiophyceae Prymnesiophyceae Imantonia rotundata group
236	Prymnesiophyceae Prymnesiophyceae Haptophyta env. 3
169	Prymnesiophyceae Prymnesiophyceae Isochrysis group
122	Prymnesiophyceae Prymnesiophyceae Haptophyta env. 5
115	Prymnesiophyceae Prymnesiophyceae Braarudosphaera group

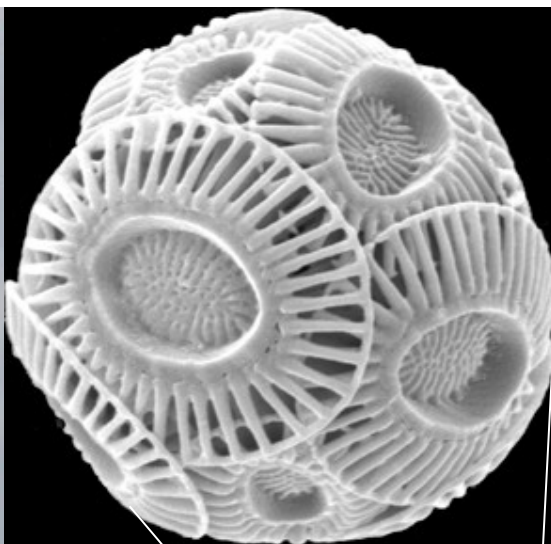
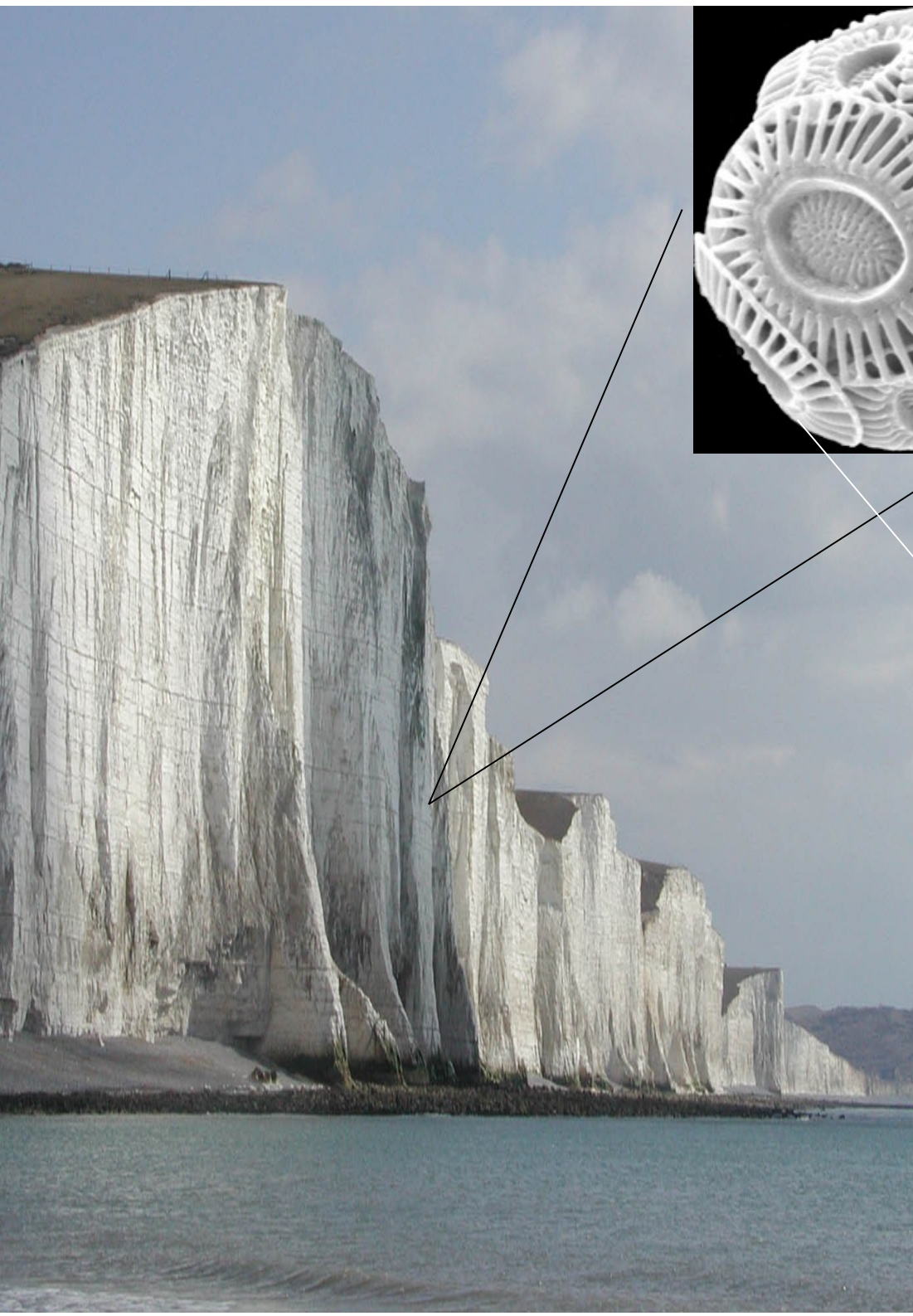


*Coccolithus pelagicus*



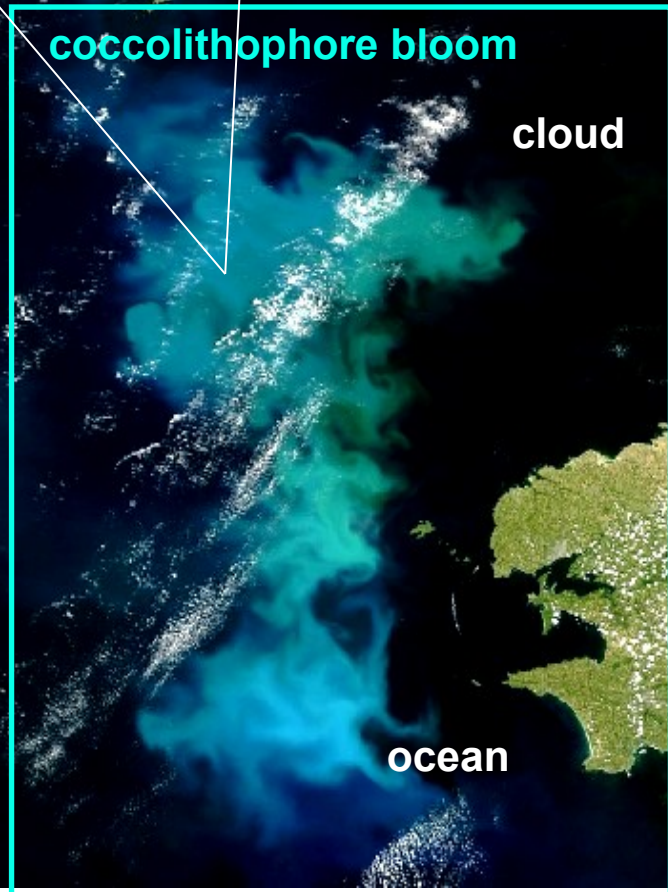
*Emiliana huxleyi*





1  $\mu$ m  
—  
10 km

CaCO<sub>3</sub>



coccolithophore bloom

cloud

**X** Roscoff

ocean

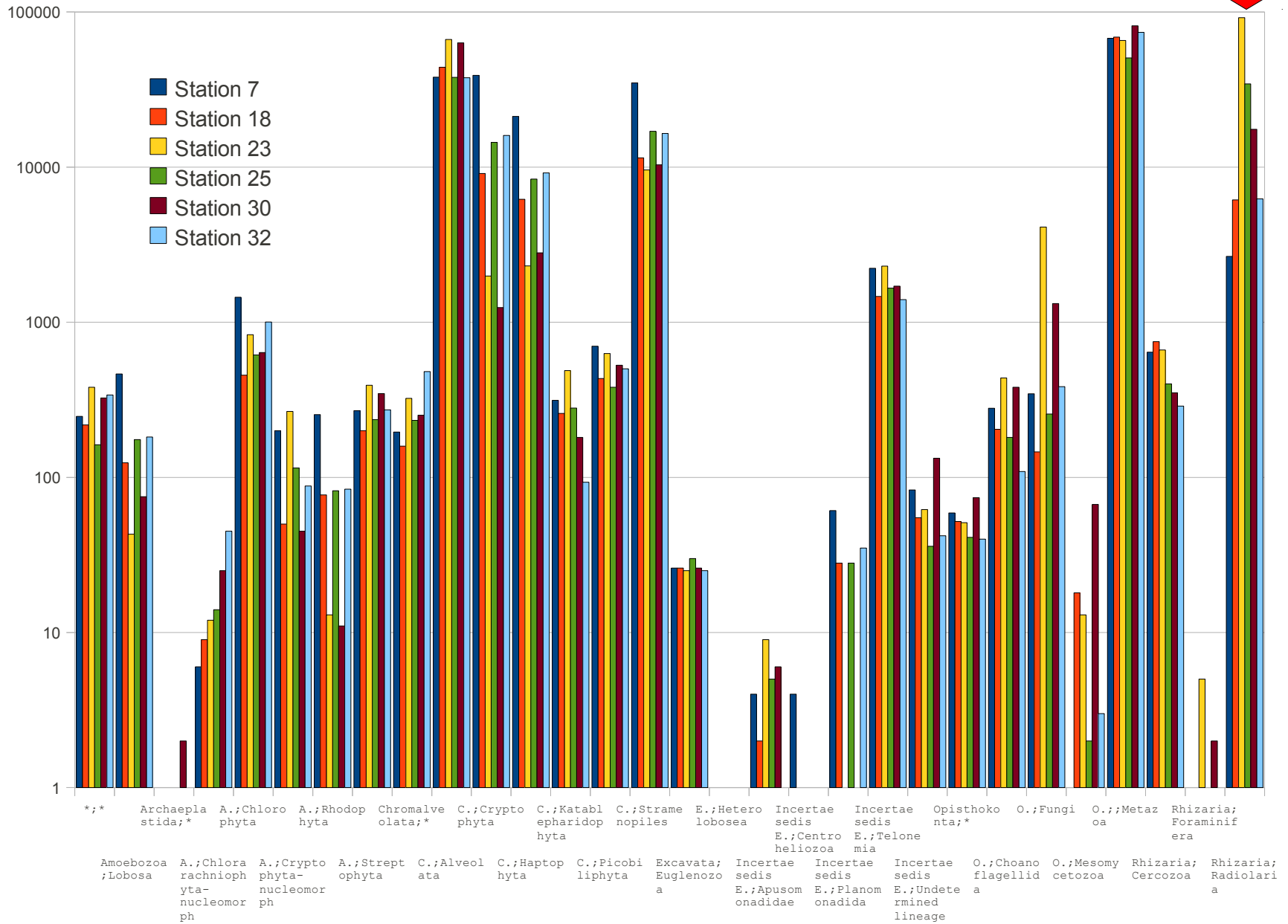
land



# Radiolaria



Repartition of assigned sequences per division

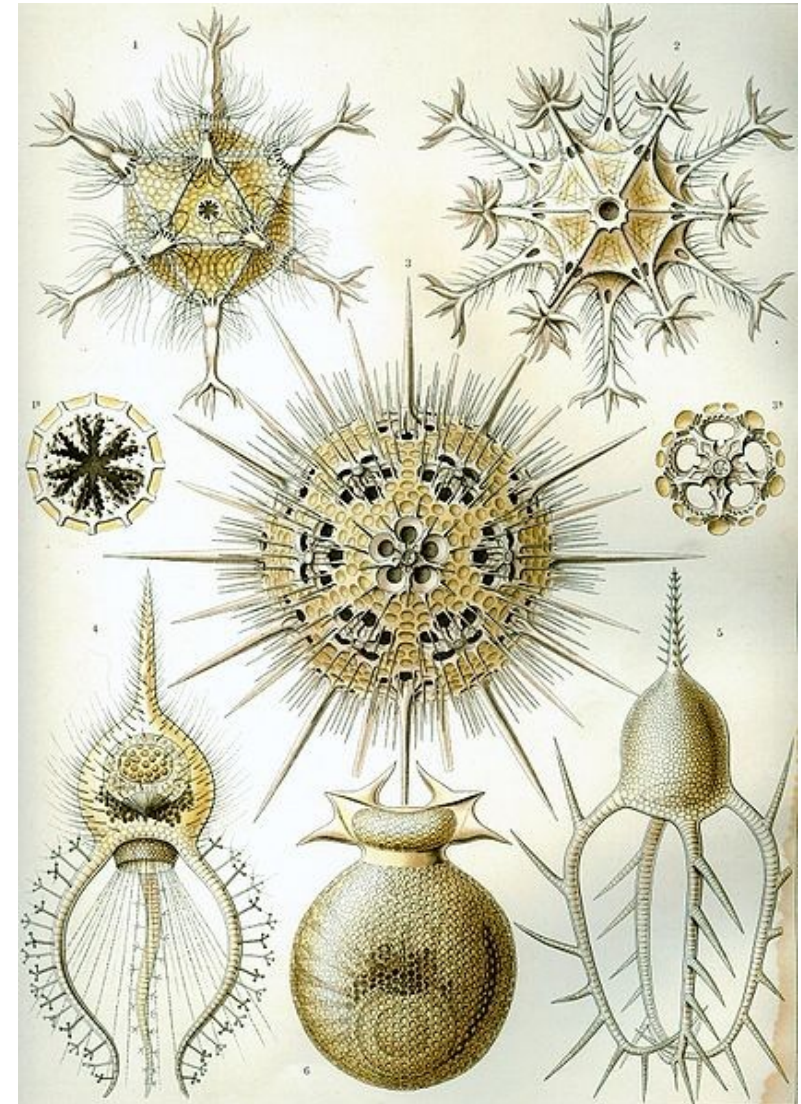
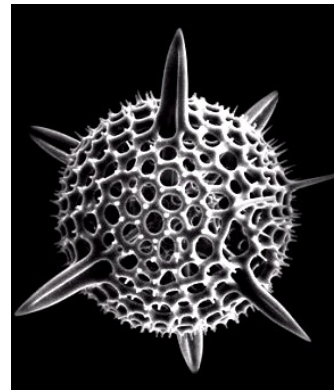
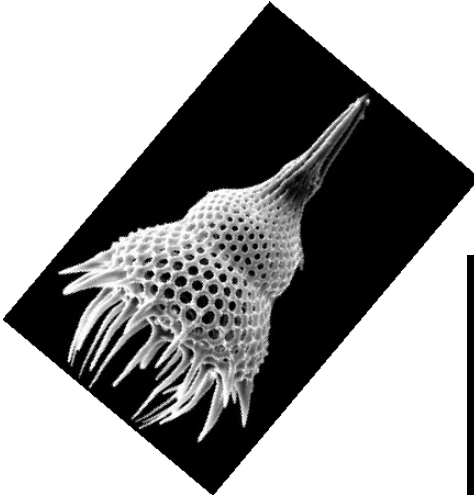
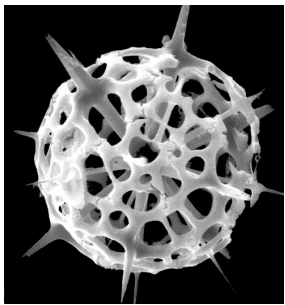


102088	Polycystinea Collodaria-Nassellarida
37543	Polycystinea Polycystinea
7022	RAD B RAD B
5094	Acantharea Acantharea
3778	RAD A RAD A
2912	Polycystinea Spumellarida
318	RAD C RAD C
82	Polycystinea *

Kunstformen der Natur (1900), Ernst Haeckel

Polycystinea -> siliceous skeleton

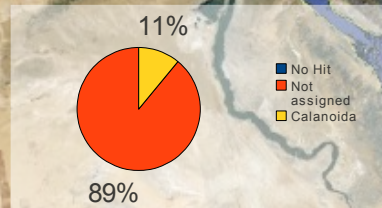
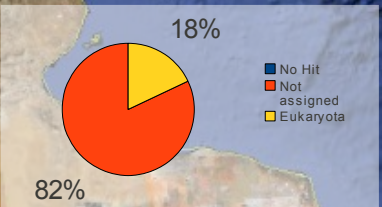
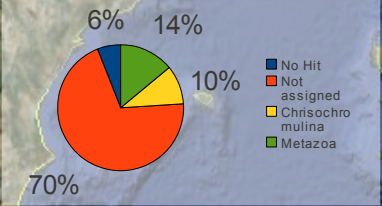
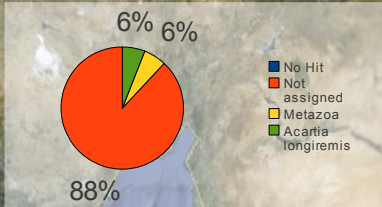
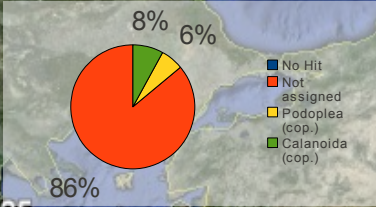
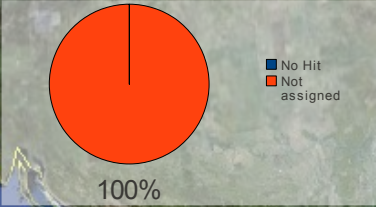
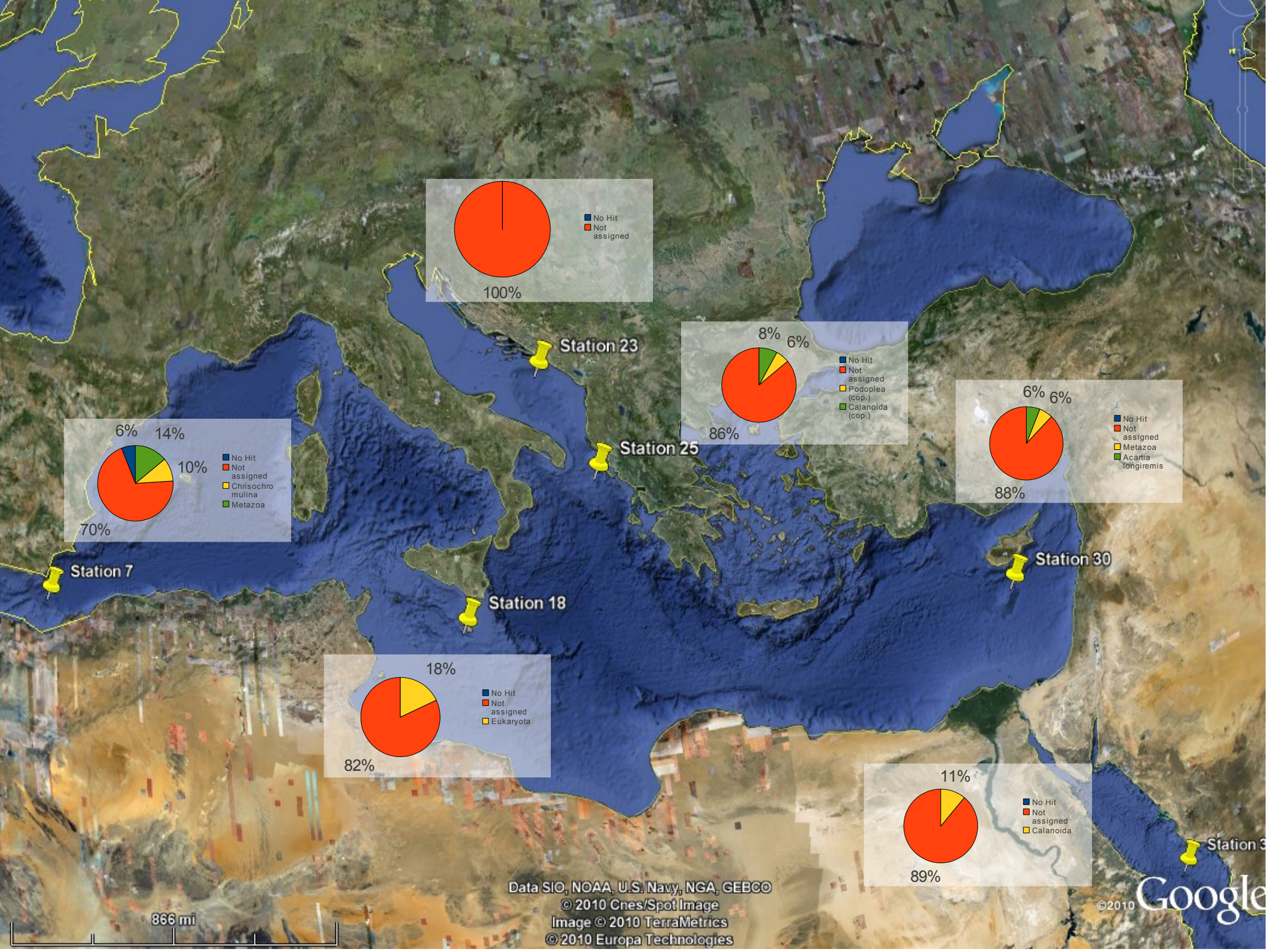
Acantharea->strontium sulfate skeleton



# Discovering new species and/or improving reference databases

- Strategy:
  - For each Station, we extract the 100 more abundant sequences not assigned by our clustering based assignation method.
    - 1) Blastn search against nucleotide database (ncbi nt)
    - 2) Assignation with Megan (Metagenome Analysis Software )





866 mi

Data SIO, NOAA, U.S. Navy, NGA, GEBCO  
© 2010 Cnes/Spot Image  
Image © 2010 TerraMetrics  
© 2010 Europa Technologies

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[Species](#) > [Copepods](#) > [Calanoida](#) > [Acartiidae](#) > [Acartia longiremis](#)

## *Acartia longiremis* (Lilljeborg, 1853)

**A common surface-dwelling copepod of the subarctic Pacific**

### Size

- Juveniles: start at 0.2mm
- Male: 0.90-1.00 mm
- Female: 0.97-1.11 mm

### Color

- Transparent, dark grey if eggs present

### Habitat

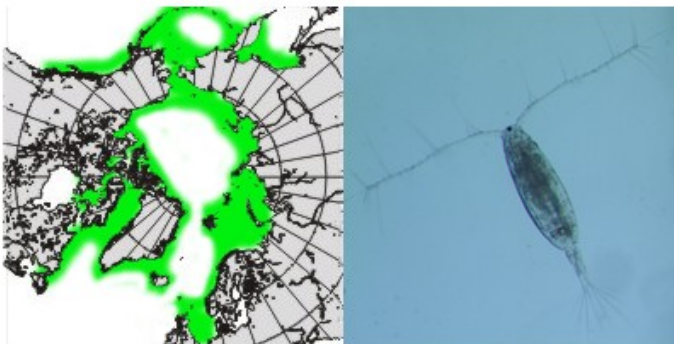
- Epipelagic (surface waters)
- Estuarine, coastal
- Arctic and Subarctic

### Feeding

- Filter-feeder
- Micro-carnivore

### Life cycle

- Female spawns repeatedly
- Small eggs scattered into water
- Generation time (egg to adult): months
- Multiple generations per year
- Present spring through late fall
- Eggs laid in fall overwinter in sediments and hatch in spring

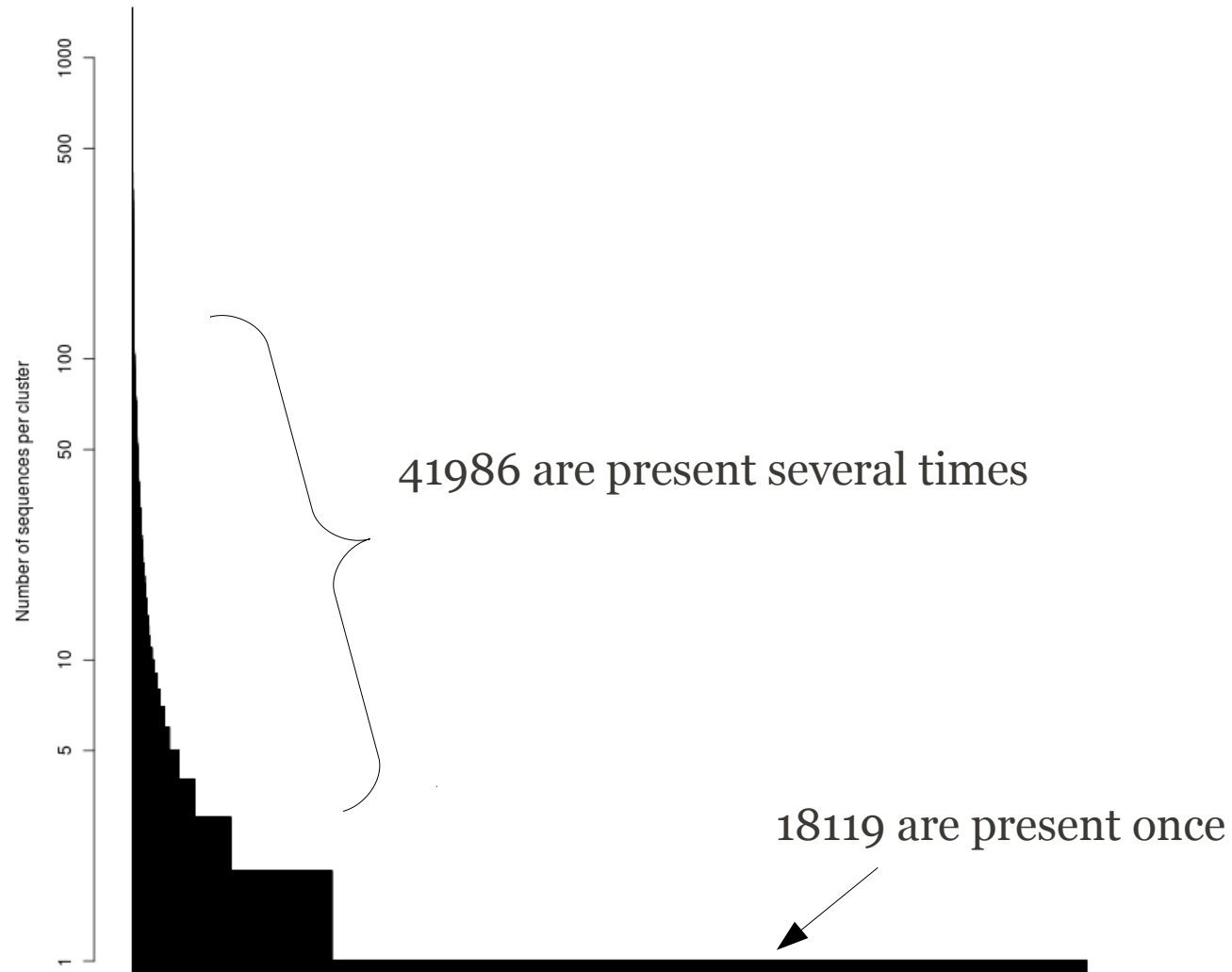


[Distribution](#)

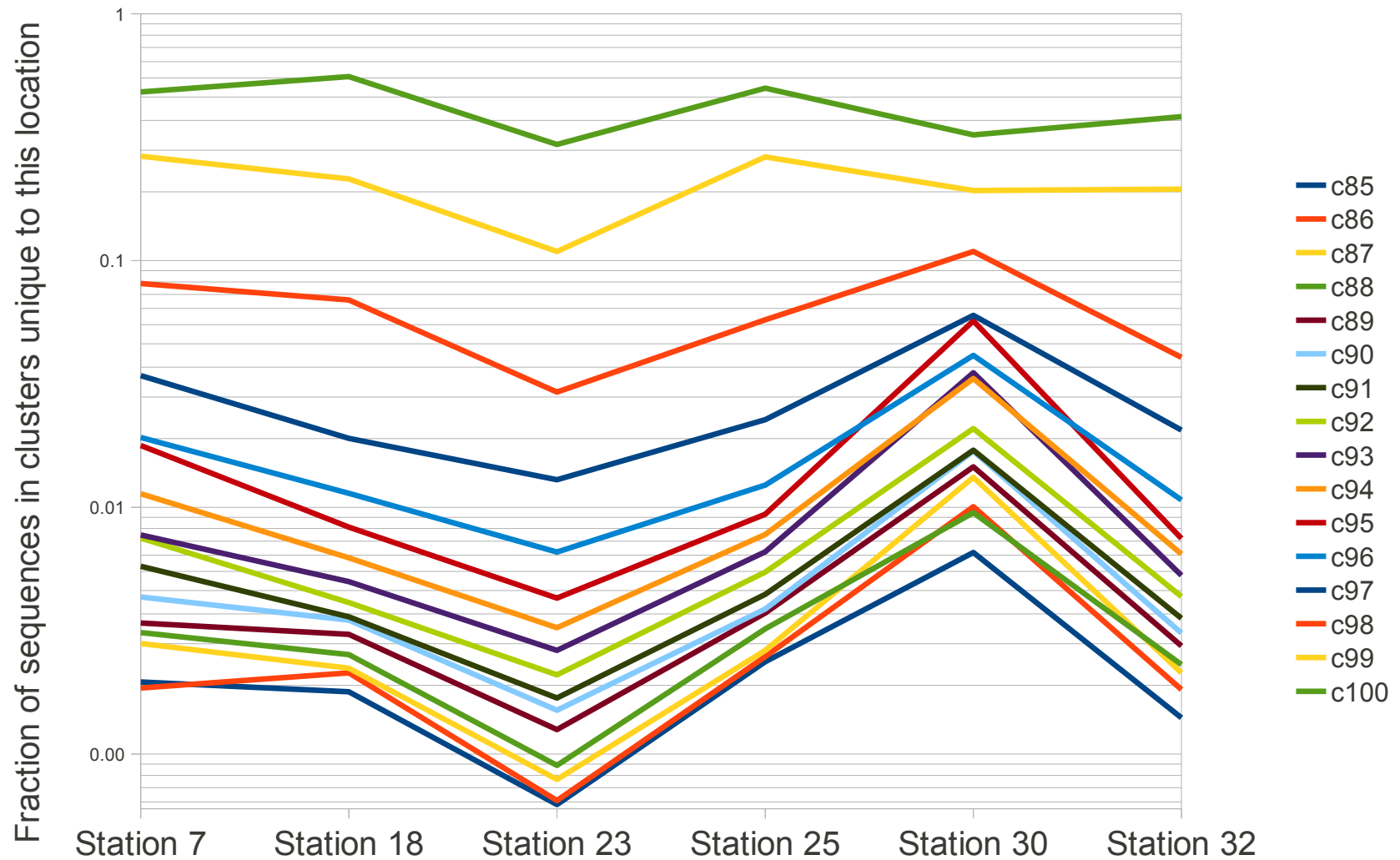
[More Images](#)

# What are unassigned sequences ?

Station 7 Unassigned sequences

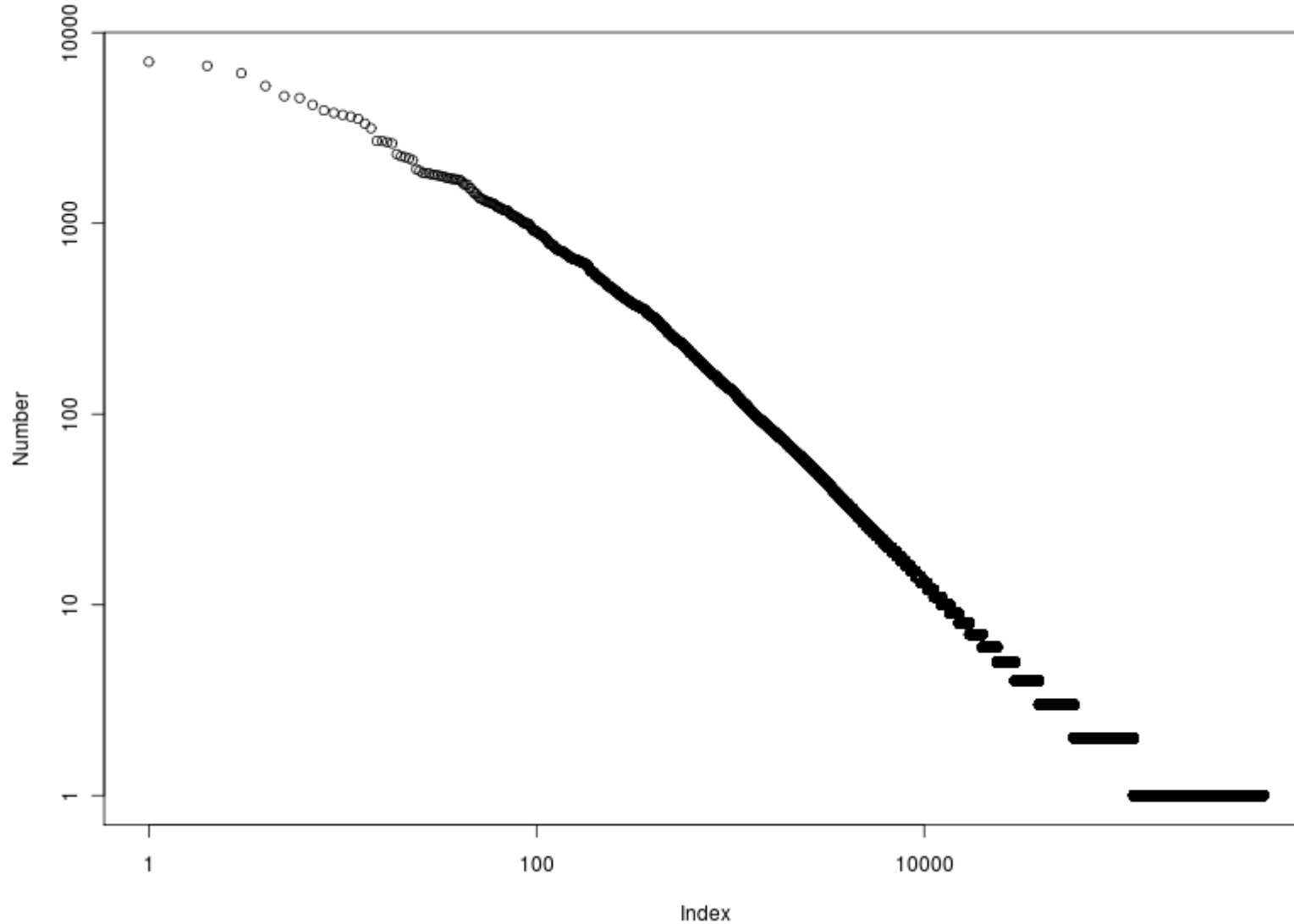


# Large overlap between stations: not so many specific clusters



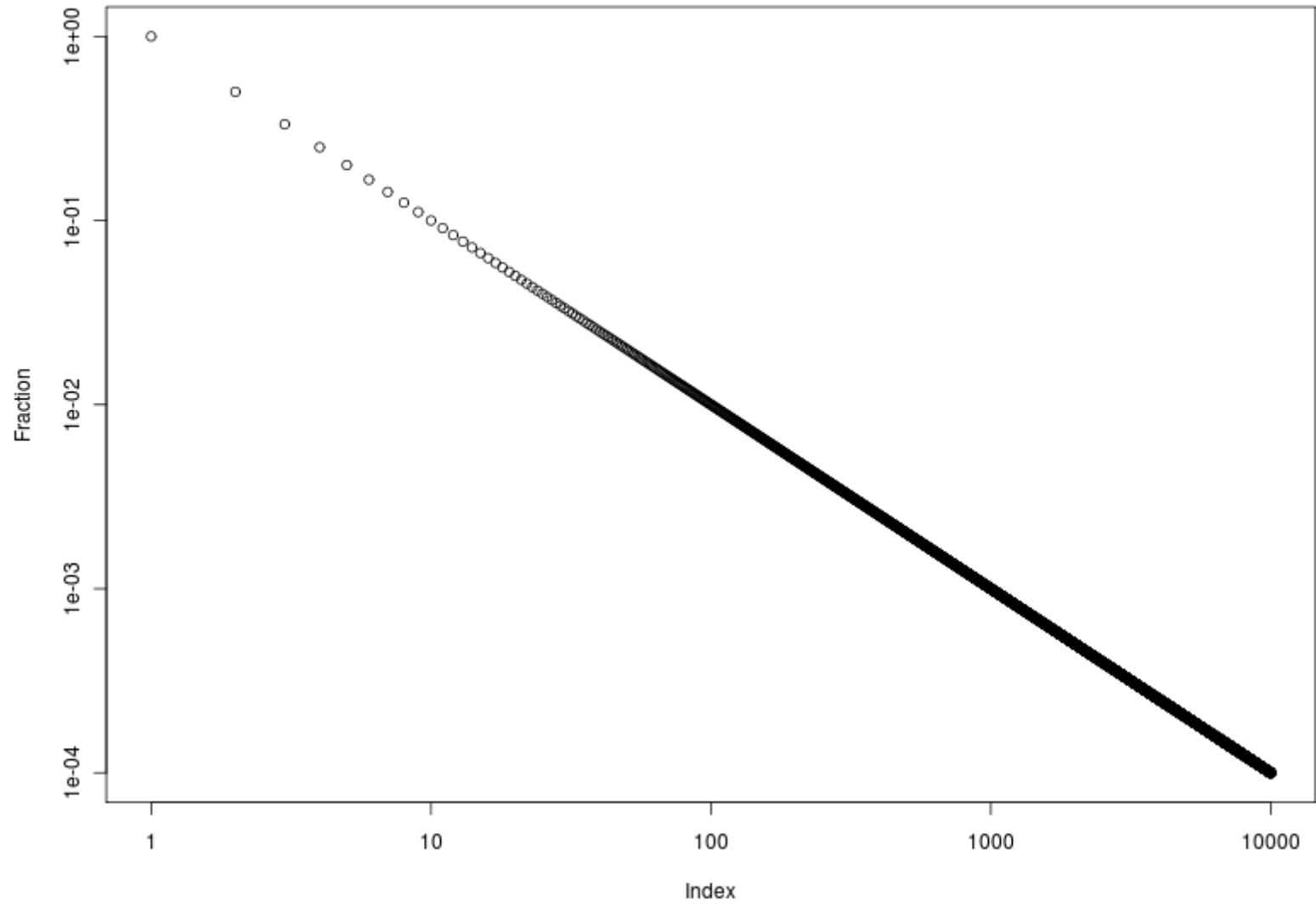
# Number of Appearance per sequence in Real Tara V9 Barcode Data

Distribution of the number of appearance per sequence

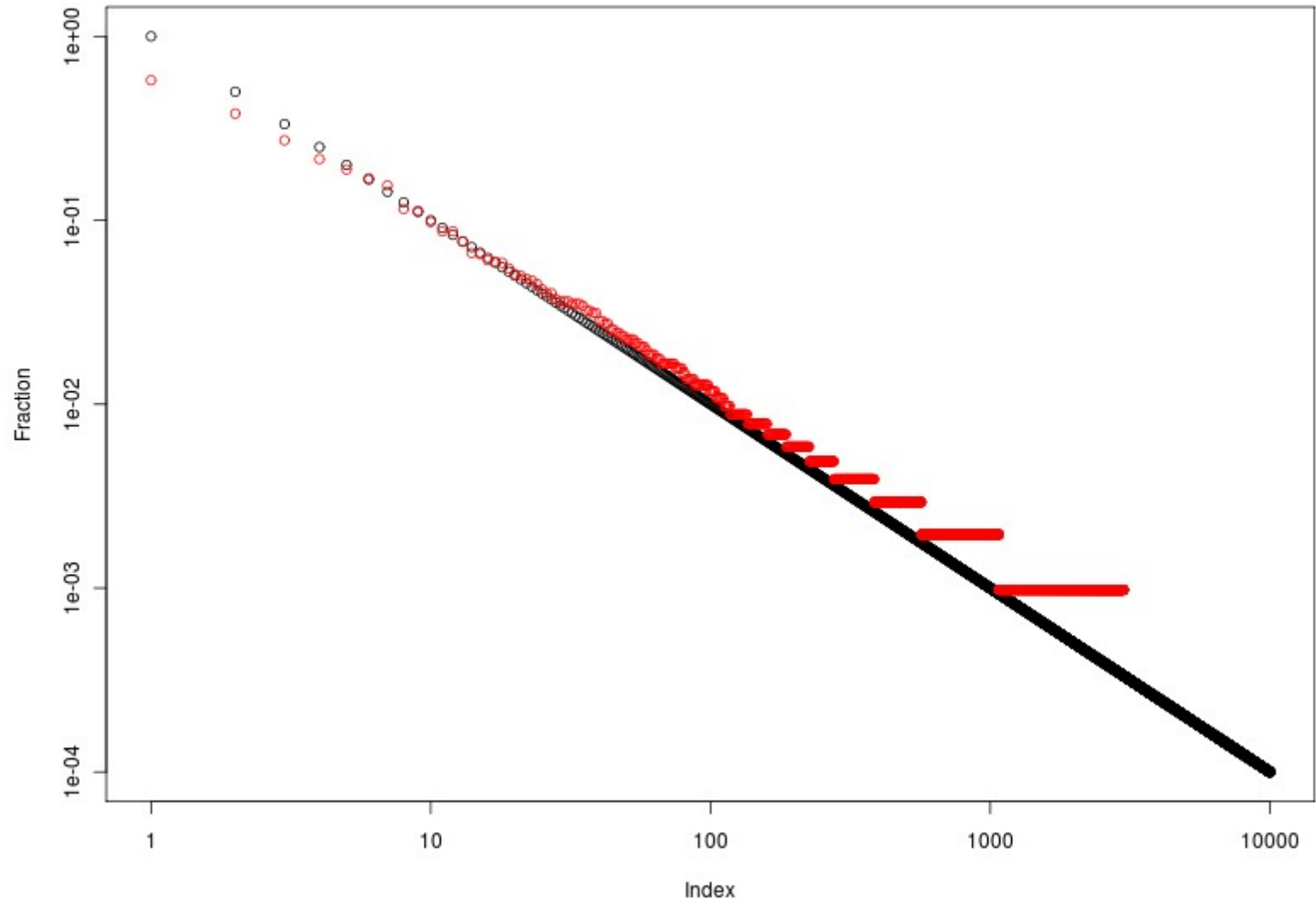




### Why Do we have many singletons?



### Why Do we have many singletons?



# Thanks

- Team Evolution du Plancton et PaléOcéan:  
(resp. Colomban de Vargas):

Fabrice Not , Lucie Bittner, Frédéric Mahé, Christophe Boutte,  
Nathalie Babic, Sarah Romac, Sébastien Colin, El Mahdi Bendif,  
Johan Decelle, Noan Le Bescot, Céline Dimier

- Adaptation et Diversité en Milieu Marin,  
UMR7144, (resp. Francois Lallier)
- Station Biologique de Roscoff (resp. Bernard  
Kloareg)